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(54) Title: TRANSGENIC PLANTS EXPRESSING PHOTORHABDUS TOXIN

(57) Abstract: Novel polynucleotide sequences that encode insect toxins TcdA and TcbA have base compositions that differ substantially from the native genes, making them more similar to plant genes. The new sequences are suitable for use for high expression in both monocots and dicots. Transgenic plants with a genome comprising a nucleic acid of SEQ ID NO: 3 or SEQ ID NO:4 are insect resistant.

TRANSGENIC PLANTS EXPRESSING PHOTORHABDUS TOXIN

## BACKGROUND OF THE INVENTION

As reported in WO98/08932, protein toxins from the genus *Photorhabdus* have been shown to have oral toxicity against insects. The toxin complex produced by *Photorhabdus luminescens* (W-14), for example, has been shown to contain ten to fourteen proteins, and it is known that these are produced by expression of genes from four distinct genomic regions: *tca*, *tcb*, *tcc*, and *tcd*. WO98/08932 discloses nucleotide sequences for the native toxin genes.

Of the separate toxins isolated from *Photorhabdus luminescens* (W-14), those designated Toxin A and Toxin B are especially potent against target insect species of interest, for example corn rootworm. Toxin A is comprised of two different subunits. The native gene *tcdA* (SEQ ID NO:1) encodes protoxin TcdA (see SEQ ID NO:1). As determined by mass spectrometry, TcdA is processed by one or more proteases to provide Toxin A. More specifically, TcdA is an approximately 282.9 kDA protein (2516 aa) that is processed to provide TcdAii, an approximately 208.2 kDA (1849 aa) protein encoded by nucleotides 265-5811 of SEQ ID NO:1, and TcdAiii, an approximately 63.5 kDA (579 aa) protein encoded by nucleotides 5812-7551 of SEQ ID NO:1.

Toxin B is similarly comprised of two different subunits. The native gene *tcbA* (SEQ ID NO:2) encodes protoxin TcbA (see SEQ ID NO:2). As determined by mass spectrometry, TcbA is processed by one or more proteases to provide Toxin B. More specifically, TcbA is an approximately 280.6 kDA (2504 aa) protein that is processed to provide TcbAii, an approximately 207.7 kDA (1844 aa) protein encoded by nucleotides 262-5793 of SEQ ID NO:2 and TcbAiii, an approximately 62.9 kDA (573 aa) protein encoded by nucleotides 5794-7512 of SEQ ID NO:2.

The native *tcdA* and *tcbA* genes are not well suited for high level expression in plants. They encode multiple destabilization sequences, mRNA splice sites, polyA addition sites and other possibly detrimental sequence motifs. In addition, the codon compositions are not like those of plant genes. WO98/08932 gives general guidance on how the toxin genes could be reengineered to more efficiently expressed in the cytoplasm of plants, and describes how plants can be transformed to incorporate the *Photorhabdus* toxin genes into their genomes.

#### SUMMARY OF THE INVENTION

In a preferred embodiment, the invention provides novel polynucleotide sequences that encode TcdA and TcbA. The novel sequences have base compositions that differ substantially from the native genes, making them more similar to plant genes. The new sequences are suitable for use for high expression in both monocots and dicots, and this feature is designated by referring to the sequences as the "hemicot" criteria, which is set forth in detail hereinafter. Other important features of the sequences are that potentially deleterious sequences have been eliminated, and unique restriction sites have been built in to enable adding or changing expression elements, organellar targeting signals, engineered protease sites and the like, if desired.

In a particularly preferred embodiment, the invention provides polynucleotide sequences that satisfy hemicot criteria and that comprise a sequence encoding an endoplasmic reticulum signal or similar targeting sequence for a cellular organelle in combination with a sequence encoding TcdA or TdbA.

More broadly, the invention provides engineered nucleic acids encoding functional *Photorhabdus* toxins wherein the sequences satisfy hemicot criteria.

The invention also provides transgenic plants with genomes comprising a novel sequence of the invention that imparts functional activity against insects.

5 BRIEF DESCRIPTION OF SEQUENCES

SEQ ID NO:1 is the native *tcdA* DNA sequence together with the corresponding encoded amino acid sequence for TcdA.

10 SEQ ID NO:2 is the native *tcbA* DNA sequence together with the corresponding encoded amino acid sequence for TcbA.

SEQ ID NO:3 is an artificial sequence encoding TcdA that is suitable for expression in monocot and dicot plants.

15 SEQ ID NO:4 is an artificial sequence encoding TdbA that is suitable for expression in monocot and dicot plants.

20 SEQ ID NO:5 is an artificial hemicot sequence that encodes the 21 amino acid ER signal peptide of 15 kDa zein from Black Mexican Sweet maize.

SEQ ID NO:6 is an artificial hemicot sequence that encodes for the full-length native TcdA protein (amino acids 22-2537) fused to the modified 15 kDa zein endoplasmic reticulum signal peptide (amino acids 1-21).

25 DETAILED DESCRIPTION

The native *Photorhabdus* toxins are protein complexes that are produced and secreted by growing bacteria cells of the genus *Photorhabdus*. Of particular interest are the proteins produced by the species *Photorhabdus*  
30 *luminescens*. The protein complexes have a molecular size of approximately 1,000 kDa and can be separated by SDS-PAGE gel analysis into numerous component proteins. The toxins contain no hemolysin, lipase, type C phospholipase, or nuclease activities. The toxins  
35 exhibit significant toxicity upon ingestion by a number of insects.



A unique feature of *Photorhabdus* is its bioluminescence. *Photorhabdus* may be isolated from a variety of sources. One such source is nematodes, more particularly nematodes of the genus *Heterorhabditis*.

5 Another such source is from human clinical samples from wounds, see Farmer et al. 1989 J. Clin. Microbiol. 27 pp. 1594-1600. These saprophytic strains are deposited in the American Type Culture Collection (Rockville, MD) ATCC #s 43948, 43949, 43950, 43951, and 43952, and are  
10 incorporated herein by reference. It is possible that other sources could harbor *Photorhabdus* bacteria that produce insecticidal toxins. Such sources in the environment could be either terrestrial or aquatic based.

The genus *Photorhabdus* is taxonomically defined as a  
15 member of the Family *Enterobacteriaceae*, although it has certain traits atypical of this family. For example, strains of this genus are nitrate reduction negative, yellow and red pigment producing and bioluminescent. This latter trait is otherwise unknown within the  
20 *Enterobacteriaceae*. *Photorhabdus* has only recently been described as a genus separate from the *Xenorhabdus* (Boemare et al., 1993 Int. J. Syst. Bacteriol. 43, 249-255). This differentiation is based on DNA-DNA hybridization studies, phenotypic differences (e.g.,  
25 presence (*Photorhabdus*) or absence (*Xenorhabdus*) of catalase and bioluminescence) and the Family of the nematode host (*Xenorhabdus*; *Steinernematidae*, *Photorhabdus*; *Heterorhabditidae*). Comparative, cellular fatty-acid analyses (Janse et al. 1990, Lett. Appl. Microbiol 10, 131-135; Suzuki et al. 1990, J. Gen. Appl. Microbiol., 36, 393-401) support the separation of  
30 *Photorhabdus* from *Xenorhabdus*.

Currently, the bacterial genus *Photorhabdus* is comprised of a single defined species, *Photorhabdus*  
35 *luminescens* (ATCC Type strain #29999, Poinar et al., 1977, Nematologica 23, 97-102). A variety of related

strains have been described in the literature (e.g., Akhurst et al. 1988 J. Gen. Microbiol., 134, 1835-1845; Boemare et al. 1993 Int. J. Syst. Bacteriol. 43 pp. 249-255; Putz et al. 1990, Appl. Environ. Microbiol., 56, 5 181-186).

The following toxin producing *Photorhabdus* strains have been deposited:

strain	accession number	date of deposit
W-14	ATCC 55397	March 5, 1993
WX1	NRRL B-21710	April 29, 1997
WX2	NRRL B-21711	April 29, 1997
WX3	NRRL B-21712	April 29, 1997
WX4	NRRL B-21713	April 29, 1997
WX5	NRRL B-21714	April 29, 1997
WX6	NRRL B-21715	April 29, 1997
WX7	NRRL B-21716	April 29, 1997
WX8	NRRL B-21717	April 29, 1997
WX9	NRRL B-21718	April 29, 1997
WX10	NRRL B-21719	April 29, 1997
WX11	NRRL B-21720	April 29, 1997
WX12	NRRL B-21721	April 29, 1997
WX14	NRRL B-21722	April 29, 1997
WX15	NRRL B-21723	April 29, 1997
H9	NRRL B-21727	April 29, 1997
Hb	NRRL B-21726	April 29, 1997
Hm	NRRL B-21725	April 29, 1997
HP88	NRRL B-21724	April 29, 1997
NC-1	NRRL B-21728	April 29, 1997
W30	NRRL B-21729	April 29, 1997
WIR	NRRL B-21730	April 29, 1997
B2	NRRL B-21731	April 29, 1997
ATCC 43948	ATCC 55878	November 5, 1996
ATCC 43949	ATCC 55879	November 5, 1996
ATCC 43950	ATCC 55880	November 5, 1996
ATCC 53951	ATCC 55881	November 5, 1996
ATCC 43952	ATCC 55882	November 5, 1996
DEPI	NRRL B-21707	April 29, 1997
DEP2	NRRL B-21708	April 29, 1997
DEP3	NRRL B-21709	April 29, 1997
P. zealandrica	NRRL B-21683	April 29, 1997
P. hepialus	NRRL B-21684	April 29, 1997
HB-Arg	NRRL B-21685	April 29, 1997
HB Oswego	NRRL B-21686	April 29, 1997
Hb Lewiston	NRRL B-21687	April 29, 1997
K-122	NRRL B-21688	April 29, 1997
HMGD	NRRL B-21689	April 29, 1997
Indicus	NRRL B-21690	April 29, 1997
GD	NRRL B-21691	April 29, 1997
PWH-5	NRRL B-21692	April 29, 1997
Megidis	NRRL B-21693	April 29, 1997
HF-85	NRRL B-21694	April 29, 1997
A. Cows	NRRL B-21695	April 29, 1997
MP1	NRRL B-21696	April 29, 1997
MP2	NRRL B-21697	April 29, 1997
MP3	NRRL B-21698	April 29, 1997
MP4	NRRL B-21699	April 29, 1997
MP5	NRRL B-21700	April 29, 1997
GL98	NRRL B-21701	April 29, 1997
GL101	NRRL B-21702	April 29, 1997
GL138	NRRL B-21703	April 29, 1997
GL155	NRRL B-21704	April 29, 1997
GL217	NRRL B-21705	April 29, 1997
GL257	NRRL B-21706	April 29, 1997

All strains were deposited in accordance with the terms of the Budapest Treaty. Strains having

accession numbers prefaced by "ATTC" were deposited on the indicated date in the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852 USA. Strains prefaced by "NRRL" were

5 deposited on the indicated date in the Agricultural Research Service Patent Culture Collection (NRRL), National Center for Agricultural Utilization Research, ARS-USDA, 1815 North University St., Peoria IL 61604 USA.

10 The present invention provides hemicot nucleic acid sequences encoding toxins from any *Photorhabdus* species or strain that produces a toxin having functional activity. Hemicot nucleic acid sequences encoding proteins homologous to such toxins are also encompassed  
15 by the invention.

Several terms that are used herein have a particular meaning and are defined as follows:

By "functional activity" it is meant herein that the protein toxins) function as insect control agents in that  
20 the proteins are orally active, or have a toxic effect, or are able to disrupt or deter feeding, which may or may not cause death of the insect. When an insect comes into contact with an effective amount of toxin delivered via transgenic plant expression, formulated protein  
25 compositions), sprayable protein compositions), a bait matrix or other delivery system, the results are typically death of the insect, or the insects do not feed upon the source which makes the toxins available to the insects.

30 By "homolog" it is meant an amino acid sequence that is identified as possessing homology to a reference *Photorhabdus* toxin polypeptide amino acid sequence.

By "homology" it is meant an amino acid sequence that has a similarity index of at least 33% and/or an  
35 identity index of at least 26% to a reference *Photorhabdus* toxin polypeptide amino acid sequence, as

scored by the GAP algorithm using the B10sum 62 protein scoring matrix Wisconsin Package Version 9.0, Genetics Computer Group GCG), Madison, WI).

By "identity" is meant an amino acid sequence that  
5 contains an identical residue at a given position,  
following alignment with a reference *Photrhabdus* toxin  
polypeptide amino acid sequence by the GAP algorithm.

By the use of the term "*Photorhabdus* toxin" it is  
meant any protein produced by a *Photorhabdus*  
10 microorganism strain which has functional activity  
against insects, where the *Photorhabdus* toxin could be  
formulated as a sprayable composition, expressed by a  
transgenic plant, formulated as a bait matrix, delivered  
via baculovirus, or delivered by any other applicable  
15 host or delivery system.

By the use of the term "toxic" or "toxicity" as used  
herein it is meant that the toxins produced by  
*Photorhabdus* have "functional activity" as defined  
herein.

By "substantial sequence homology" is meant either:  
20 a DNA fragment having a nucleotide sequence sufficiently  
similar to another DNA fragment to produce a protein  
having similar biochemical properties; or a polypeptide  
having an amino acid sequence sufficiently similar to  
25 another polypeptide to exhibit similar biochemical  
properties.

As with other bacterial toxins, the rate of mutation  
of the bacteria in a population causes many related  
toxins slightly different in sequence to exist. Toxins  
30 of interest here are those which produce protein  
complexes toxic to a variety of insects upon exposure, as  
described herein. Preferably, the toxins are active  
against *Lepidoptera*, *Coleoptera*, *Homoptera*, *Diptera*,  
*Hymenoptera*, *Dictyoptera* and *Acarina*. The inventions  
35 herein are intended to capture the protein toxins  
homologous to protein toxins produced by the strains

herein and any derivative strains thereof, as well as any protein toxins produced by *Photorhabdus*. These homologous proteins may differ in sequence, but do not differ in function from those toxins described herein.

5 Homologous toxins are meant to include protein complexes of between 300 kDa to 2,000 kDa and are comprised of at least two 2) subunits, where a subunit is a peptide which may or may not be the same as the other subunit. Various protein subunits have been identified and are taught in  
10 the Examples herein. Typically, the protein subunits are between about 18 kDa to about 230 kDa; between about 160 kDa to about 230 kDa; 100 kDa to 160 kDa; about 80 kDa to about 100 kDa; and about 50 kDa to about 80 kDa.

As discussed above, some *Photorhabdus* strains can be  
15 isolated from nematodes. Some nematodes, elongated cylindrical parasitic worms of the phylum *Nematoda*, have evolved an ability to exploit insect larvae as a favored growth environment. The insect larvae provide a source of food for growing nematodes and an environment in which  
20 to reproduce. One dramatic effect that follows invasion of larvae by certain nematodes is larval death. Larval death results from the presence of, in certain nematodes, bacteria that produce an insecticidal toxin which arrests larval growth and inhibits feeding activity.

25 Interestingly, it appears that each genus of insect parasitic nematode hosts a particular species of bacterium, uniquely adapted for symbiotic growth with that nematode. In the interim since this research was initiated, the name of the bacterial genus *Xenorhabdus*  
30 was reclassified into the *Xenorhabdus* and the *Photorhabdus*. Bacteria of the genus *Photorhabdus* are characterized as being symbionts of *Heterorhabditus* nematodes while *Xenorhabdus* species are symbionts of the *Steinernema* species. This change in nomenclature is  
35 reflected in this specification, but in no way should a

change in nomenclature alter the scope of the inventions described herein.

The peptides and genes that are disclosed herein are named according to the guidelines recently published in the Journal of Bacteriology "Instructions to Authors" p. i-xii Jan. 1996), which is incorporated herein by reference.

Transformation methods useful in carrying out the invention are well known, and are described, for example, in WO98/08932.

#### Hemicot tcdA and tcbA

SEQ ID NO: 3 is the nucleotide sequence for an engineered *tcdA* gene in accordance with the invention. SEQ ID NO: 4 is the nucleotide sequence for an engineered *tcbA* gene in accordance with the invention.

The following Tables 1 and 2 identify significant features of the engineered *tcdA* and *tcbA* genes.

Table 1  
*tcdA*

Feature	nucleotides of SEQ ID NO:3
<i>NcoI</i>	1-6
<i>HindIII</i>	48-53
<i>KpnI</i>	246-254
sequence encoding <i>TcbAii</i>	267-5798
<i>NheI</i>	333-338
<i>BglII</i>	1215-1220
<i>ClaI</i>	2604-2609
<i>PstI</i>	4015-4020
<i>AgeI</i>	5088-5093
<i>MunI</i>	5598-5603
<i>XbaI</i>	5778-5783
sequence encoding <i>TcbAiii</i>	5799-7517
<i>AflIII</i>	5853-5858
<i>SphI</i>	6439-6444
<i>SfuI</i>	7392-7397
<i>SacI</i>	7519-7524
<i>XhoI</i>	7522-7527
<i>StuI</i>	7528-7533
<i>NotI</i>	7533-7538

20

Table 2  
*tcbA*

Feature	nucleotides of SEQ ID NO:5
<i>NcoI</i>	1-6
<i>HindIII</i>	48-53

<i>KpnI</i>	246-251
sequence encoding <i>TcbAii</i>	267-5798
<i>NheI</i>	333-338
<i>BglII</i>	1215-1220
<i>ClaI</i>	2604-2609
<i>PstI</i>	4015-4020
<i>AgeI</i>	5088-5093
<i>MunI</i>	5598-5603
<i>XbaI</i>	5778-5783
sequence encoding <i>TcbAiii</i>	5799-7517
<i>AflIII</i>	5853-5858
<i>SphI</i>	6439-6444
<i>SfuI</i>	7392-7397
<i>SacI</i>	7519-7524
<i>SfuI</i>	7392-7397
<i>SacI</i>	7519-7524
<i>XhoI</i>	7522-7527
<i>StuI</i>	7528-7533
<i>NotI</i>	7535-7540

It should be noted that the proteins encoded by the plant-optimized *tcdA* (SEQ ID NO:3) and *tcbA* (SEQ ID NO:5) differ from the native proteins by the addition of an Ala residue at position #2. This modification was made to accommodate the *NcoI* site which spans the ATG start codon.

The following Table 3 compares the codon composition of the engineered *tcdA* gene of SEQ ID NO:3 and engineered *tcbA* gene of SEQ ID NO:5 with the codon compositions of the native genes, the typical dicot genes, and maize genes.

Table 3

amino acid	codon	% in SEQ ID NO:3	% in <i>tcdA</i>	% in SEQ ID NO:5	% in <i>tcbA</i>	% in dicot	% in maize
Ala	GCT	62	21	69	41	42	24
	GCC	26	32	27	17	27	34
	GCA	11	25	4	22	25	18
	GCG	0	21	0	21	6	24
Arg	AGG	48	0	60	2	25	26
	CGC	22	36	18	16	11	24
	AGA	20	11	15	6	30	15
	CGT	11	39	7	57	21	11
	CGG	0	7	0	13	4	15
	CGA	0	8	0	6	8	9
Asn	AAC	100	32	100	33	55	68
	AAT	0	68	0	67	45	32
Asp	GAC	67	22	70	25	42	63



amino acid	codon	% in SEQ ID NO:3	% in tcdA	% in SEQ ID NO:5	% in tcbA	% in dicot	% in maize
	GAT	33	78	30	75	58	37
Cys	TGC	100	30	100	19	56	68
	TGT	0	70	0	81	44	32
End	TGA	100	0	100	0	33	59
	TAG	0	0	0	0	19	21
	TAA	0	100	0	100	48	20
Gln	CAA	65	61	74	53	59	38
	CAG	35	39	26	47	41	62
Glu	GAG	100	24	98	36	51	71
	GAA	0	76	2	64	49	29
Gly	GGT	67	37	64	44	33	20
	GGC	32	36	36	22	16	42
	GGA	1	20	0	19	38	19
	GGG	0	8	0	16	12	20
His	CAC	62	40	72	31	46	62
	CAT	38	60	28	69	54	38
Ile	ATC	73	34	65	24	37	58
	ATT	27	51	35	59	45	28
	ATA	0	15	0	17	18	14
Leu	CTC	54	11	59	7	28	26
	TTG	29	17	25	32	26	15
	CTT	16	9	15	7	19	17
	TTA	0	18	0	19	10	5
	CTG	0	32	0	29	9	29
	CTA	0	13	0	7	8	8
Lys	AAG	99	79	99	75	61	78
	AAA	1	21	1	25	39	22
Met	ATG	100	100	100	100	100	100
Phe	TTC	100	42	100	41	55	71
	TTT	0	58	0	59	45	29
Pro	CCA	74	30	91	26	42	26
	CCT	22	28	7	20	32	22
	CCC	4	14	3	7	17	24
	CCG	0	27	0	47	9	28
Ser	TCC	47	19	55	11	18	23
	TCT	35	15	30	15	25	15
	AGC	18	22	15	18	18	23
	AGT	0	20	0	31	14	9
	TCG	0	7	0	8	6	14
	TCA	0	17	0	17	19	16
Thr	ACC	60	41	64	31	30	37
	ACT	28	25	32	34	35	20
	ACA	12	21	4	18	27	21
	ACG	0	13	0	18	8	22
Trp	TGG	100	100	100	100	100	100
Tyr	TAC	100	24	100	19	57	73
	TAT	0	76	0	81	43	27
Val	GTC	69	27	73	11	20	31
	GTG	21	17	22	27	29	39
	GTT	10	34	3	48	39	21
	GTA	0	22	2	14	12	8

## EXAMPLE 1

Design Of Plant Codon-Biased Genes Encoding W-14 Peptides  
TcbA and TcdA

5

## A. Gene Design

The coding strands of the native DNA sequences of the *Photorhabdus* W-14 genes encoding peptides TcbA and TcdA were scanned for the presence of deleterious sequences such as the Shaw/Kamen RNA destabilizing motif ATTTA, intron splice recognition sites, and poly A addition motifs. This was done using the MacVector Sequence Analysis Software (Oxford Molecular Biology Group, Symantec Corp.), using a custom Nucleic Acid Subsequence File. The native sequence was also searched for runs of 4 or more of the same base.

Motif searching of the native W-14 *tcbA* and *tcdA* genes revealed the presence of many potentially deleterious sequences in the protein coding strands, as summarized in Table 4. Not shown, but also present, were many runs of four or more single residues (e.g. the native *tcbA* gene has 81 runs of four A's).

Table 4

Native Gene	ATTTA	5' Splice	3' Splice	Poly A Addition*	RNAP II term.
<i>tcbA</i>	18	7	17	46	0
<i>tcdA</i>	18	7	13	77	1

\* Totals of 16 different motifs.

Analyses of eukaryotic genes and plant genes in particular have shown that CG & TA doublets are underrepresented, while the genes are enriched in CT & TG doublets. The sequences of the hemicot biased genes have accordingly been adjusted to encompass these base compositions and to have G+C compositions of about 53%, similar to many plant genes. When compared to the native W-14 *tcbA* and *tcdA* genes, the plant-biased genes have a much more uniform G+C distribution.

Nucleotide changes to remove potentially deleterious sequences were chosen to simultaneously adjust the codon composition of the coding region to more closely reflect that of plant genes. A framework for these changes was provided by the codon bias tables prepared for maize and dicot genes shown in Table 3.

Comparison of codon compositions of the native W-14 genes to maize and dicot genes revealed that the W-14 genes contain a very different preference set of the degenerate codons for the 18 amino acids for which there is a choice (Table 3). For each of 8 amino acids (Phe, Tyr, Cys, Arg, Asn, Lys, Glu, and Gly) in both W-14 genes, the most abundant codon is different from the preferred codons found in either maize or dicot genes. One might expect that translational difficulties would be encountered in efforts to produce in plants proteins (such as TcbA and TcdA) having high relative amounts of these amino acids from mRNAs having large numbers of nonpreferred codons. There is a marked difference in distribution of the codon compositions specifying the other 10 amino acids. For His, Gln, Ile, Val, and Asp, the dicot-preferred codons are found as the most abundant ones in both W-14 genes. For Leu, Thr, Ser, and Ala, the maize preferred codons are the most abundant codon choices found in the *tcdA* gene. In contrast, the *tcbA* gene contains only the CCG (Pro) maize-preferred codon as the highest abundance choice.

In making the codon choices, doublet contents were considered, so that adjacent codons preferably did not form CG or TA doublets (which are underrepresented in eukaryotic genes; 1, 4), while CT or TG doublets (which are enriched in eukaryotic genes ibid.) were created when possible.

Choices were also made to utilize a diversity of codons for Met, Trp, Asn, Asp, Cys, Glu, His, Ile, Lys, Phe, Thr, and Tyr.

The sequences were also designed to encode unique 6-bp recognition sites for restriction enzymes, spaced about every 1200 bp. Finally, an additional codon (GCT; Ala) was inserted at the second position to encode an Nco I recognition site encompassing the ATG (Met) start codon. Additional recognition sites were included after

the stop codon to facilitate subsequent cloning steps into expression vectors. These features are set forth above in Tables 1 and 2.

The new *tcdA* and *tcbA* genes of SEQ ID NO:3 and SEQ ID NO:4 share 73.5%, and 72.6% identity, respectively, to their native W-14 counterparts (Wisconsin Genetics Computer Group, GAP algorithm).

#### B. Gene Synthesis

The complete synthesis of the plant codon-biased *tcbA* and *tcdA* genes was performed under contract by Operon Technologies, Inc. (OPTI, Alameda, CA). Basically, chemically synthesized oligonucleotides of appropriate sequence were assembled into DNA pieces about 500 bases long. These were joined together end-to-end (presumably by means of appropriately placed restriction enzyme sites) into four larger pieces of roughly 2 kilobase pairs (kbp) each; therefore each comprised about 1/4 of the entire coding region of the particular gene. DNA sequence of the pieces was confirmed at this step. If mistakes in sequence were present, the appropriate oligonucleotides were re-synthesized, and the assembly process was repeated. Once gene fractional parts were sequence verified, they were assembled in pairs to make the gene halves, and again sequence verified. Finally, the two halves were joined, and the sequences of the junctions between the halves was verified. Therefore, each part of the new gene was sequence verified at least twice.

It should be noted that attempts to express the native *tcbA* or *tcdA* genes in standard *Escherichia coli* cloning strains suggests that production of these proteins is lethal. Lethality problems may be encountered if standard cloning vectors having leaky expression from inherent *lacZ* promoters are used to assemble these genes.

### C. Addition Of Endoplasmic Reticulum Targeting Peptide To Tcda Coding Region

It is known to those in the field of plant gene expression that proteins are specifically directed into the endoplasmic reticulum (ER) by means of a short signal peptide which is removed during or after the transport process through the ER membrane. The mature (processed) protein is incorporated into the ER endomembrane or is released into the ER lumen where the transported protein may be uniquely folded (aided by chaperonins), modified by glycosylation, accumulated in the vacuole, or additionally translocated (by secretion). These processes are reviewed by Gomord and Faye [V. Gomord and L. Faye, (1996) *Signals and mechanisms involved in intracellular transport of secreted proteins in plants*. Plant Physiol. Biochem. 34:165-181] and by Bar-Peled et al. [M. Bar-Peled, D. C. Bassham, and N. V. Raikhel, (1996) *Transport of proteins in eukaryotic cells: more questions ahead*. Plant Molec. Biology 32:223-249]. It is also known that the subcellular recognition mechanisms for an ER signal peptide are evolutionarily somewhat conserved, since the ER signal for a protein normally produced in monocot (maize) cells is recognized and processed normally by dicot (tobacco) cells. This is exemplified by the maize 15 kDa zein ER signal peptide [L. M. Hoffman, D. D. Donaldson, R. Bookland, K. Rashka, and E. M. Herman, (1987) *Synthesis and protein body deposition of maize 15-kd zein in transgenic tobacco seeds*. EMBO J. 6:3213-3221, and U.S. Patent 5589616]. Further, it is known that the ER signal peptide derived from one protein can direct the translocation of a different protein if it is appropriately attached to the second protein by genetic engineering methods [D. C. Hunt and M. J. Chrispeels, (1991) *The signal peptide of a vacuolar protein is necessary and sufficient for the efficient secretion of a cytosolic protein*. Plant

Physiol. 96:18-25, and Denecke, J., J. Botterman, and R. Deblaere (1990) *Protein secretion in plants can occur via a default pathway*. Plant Cell 2:51-59]. Therefore, one may expose a protein *in vivo* to different biochemical environments by directing its accumulation in the cytosol (by not providing a signal peptide sequence), or in the ER/vacuole (by provision of an appropriate signal peptide.)

The ER signal peptide of maize 15 kDa zein proteins is known to comprise the first 20 amino acids encoded by the zein coding region. Two examples of such signal peptides the ER signal peptide of 15 kDa zein from A5707 maize, NCBI Accession # M72708, and the ER signal peptide of 15 kDa zein from Black Mexican Sweet maize, NCBI Accession # M13507. There is only a single amino acid difference (Ser vs Cys at residue 17) between these signal peptides.

SEQ ID NO:5 is a modified sequence coding the ER signal peptide of 15 kDa zein from Black Mexican Sweet maize. The modifications embodied in this sequence were made to accommodate the different monocot/dicot codon usages and other sequence motif considerations discussed above in the design of the plant-optimized *tcdA* coding region. The sequence includes an additional Ala residue at position #2 to accommodate the *NcoI* site which spans the ATG start codon.

SEQ ID NO:6 gives a sequence coding for the full-length native TcdA protein (amino acids 22-2537) fused to the modified 15 kDa zein endoplasmic reticulum signal peptide (amino acids 1-21).

#### Example 2

Transformation Of Tobacco With *Agrobacterium* Carrying Plasmid pDAB2041 Encoding *Photobacterium* Toxins  
A. Plasmid pDAB2041

Preparation of tobacco transformation vectors was accomplished in three steps. First, a modified plant-optimized *tcdA* coding region was ligated into a tobacco

plant expression cassette plasmid. In this step, the coding region was placed under the transcriptional control of a promoter functional in tobacco plant cells. RNA transcription termination and polyadenylation were mediated by a downstream copy of the terminator region from the *Agrobacterium* nopaline synthase gene. Two plasmids designed to function in this role are pDAB1507 and pDAB2006. In the second step, the complete gene comprised of the promoter, coding region, and terminator region was ligated between the T-DNA borders of an *Agrobacterium* binary vector, pDAB1542. Also positioned between the T-DNA borders was a plant selectable marker gene to allow selection of transformed tobacco plant cells. In the third step, the engineered binary vector plasmid was conjugated from its *E. coli* host strain into a disabled *Agrobacterium tumefaciens* strain capable of transforming tobacco plant cells that regenerate into fertile transgenic plants.

It is a feature of plasmid pDAB1507 that any coding region having an *Nco*I site at its 5' end and a *Sac*I site 3' to the coding region, when cloned into the unique *Nco*I and *Sac*I sites of pDAB1507, is placed under the transcriptional control of an enhanced version of the CaMV 35S promoter. It is also a feature of pDAB1507 that the 5' untranslated leader (UTR) sequence preceding the *Nco*I site comprises a modified version of the 5' UTR of the MSV coat protein gene, into which has been cloned an internally deleted version of the maize *Adh1S* intron 1. Additionally it is a feature of pDAB1507 that transcription termination and polyadenylation of the mRNA containing the introduced coding region are mediated by termination/Poly A addition sequences derived from the nopaline synthase (Nos) gene. Finally, it is a feature of pDAB1507 that the entire assembly of promoter/coding region/3'UTR can be obtained as a single DNA fragment by cleavage at the flanking *Not*I sites.

It is a feature of plasmid pDAB2006 that any coding region having an *Nco*I site at its 5' end and a *Sac*I site 3' to the coding region, when cloned into the unique *Nco*I and *Sac*I sites of pDAB2006, is placed under the transcriptional control of the CaMV 35S promoter. It is also a feature of pDAB2006 that the 5' untranslated leader (UTR) sequence preceding the *Nco*I site comprises a polylinker. Additionally it is a feature of pDAB2006 that transcription termination and polyadenylation of the mRNA containing the introduced coding region are mediated by termination/Poly A addition sequences derived from the nopaline synthase (Nos) gene. Finally, it is a feature of pDAB2006 that the entire assembly of promoter/coding region/3'UTR can be obtained as a single DNA fragment by cleavage at the flanking *Not*I sites.

It is a feature of pDAB1542 that any DNA fragment flanked by *Not*I sites can be cloned into the unique *Not*I site of pDAB1542, thus placing the introduced fragment between the T-DNA borders, and adjacent to the neomycin phosphotransferase II (kanamycin resistance) gene.

To prepare a plant-expressible gene to produce the non-targeted TcdA protein in tobacco plant cells, DNA of a plasmid (pA0H\_4-OPTI) containing the plant-optimized *tcdA* coding region, (SEQ ID No:3) was cleaved with restriction enzymes *Nco*I and *Sac*I, and the large 7550 bp fragment was ligated to similarly-cut DNA of plasmid pDAB1507 to produce plasmid pDAB2040. DNA of pDAB2040 was then digested with *Not*I, and the 8884 bp fragment was ligated to *Not*I digested DNA of pDAB1542 to produce plasmid pDAB2041. This plasmid was then conjugated by triparental mating [Firoozabady, E., D. L. DeBoer, D. J. Merlo, E. L. Halk, L. N. Amerson, K. E. Rashka, and E. E. Murray (1987) *Transformation of cotton (Gossypium hirsutum L.) by Agrobacterium tumefaciens and regeneration of transgenic plants*. Plant Molec. Biol.



10:105-116] from the host *Escherichia coli* strain (XL1-Blue, Stratagene, La Jolla, CA), into the nontumorigenic *Agrobacterium tumefaciens* strain EHA101S, which is a spontaneous streptomycin-resistant mutant of strain  
5 EHA101 (Hood, E. E., G. L. Helmer, R. T. Fraley, and M.-D. Chilton (1986) *The hypervirulence of Agrobacterium tumefaciens A281 is encoded in a region of pTiBo542 outside of T-DNA*. J. Bacteriol. 168:1291-1301). Strain EHA101S(pDAB2041) was then used to produce transgenic  
10 tobacco plants that expressed the TcdA protein.

B. Plasmid pRK2013

To prepare a plant-expressible gene to produce the endoplasmic reticulum-targeted TcdA protein in tobacco plant cells, DNA of a plasmid (pA0H\_4-ER) containing the  
15 plant-optimized, ER-targeted *tcdA* coding region, (SEQ ID No:6) was cleaved with restriction enzymes *NcoI* and *SacI*, and the large 7610 bp fragment was ligated to similarly-cut DNA of plasmid pDAB2006 to produce plasmid pDAB1833. DNA of pDAB1833 was then digested with *NotI*, and the 8822  
20 bp fragment was ligated to *NotI* digested DNA of pDAB1542 to produce plasmid pDAB2052. This plasmid was then conjugated by triparental mating from the host *Escherichia coli* strain (XL1-Blue), into the nontumorigenic *Agrobacterium tumefaciens* strain EHA101S.  
25 Strain EHA101S(pDAB2052) was then used to produce transgenic tobacco plants that expressed the TcdA protein containing an amino terminus endoplasmic reticulum targeting peptide.

30 C. Transfer of Plasmid pDAB2041 Into *Agrobacterium* Strain EHA101S

Cultures of *E. coli* carrying the engineered Ti plasmid pDAB2041 (plasmid containing the rebuilt Toxin A gene, *tcdA*), *E. coli* carrying the plasmid pRK2013, and  
35 *Agrobacterium* strain EHA101S were grown overnight, then mixed 1:1:1 on plain LB medium solidified with agar and

cultured in the dark at 28°C. Two days later, the lawn of bacteria was scraped up with a loop, suspended in plain LB medium, vortexed, and then diluted 1:10<sup>4</sup>, 1:10<sup>5</sup>, and 1:10<sup>6</sup> fold in plain LB liquid medium. Aliquots of these dilutions were spread on selective plates containing medium YEP plus erythromycin (100 mg/L) and streptomycin (250 mg/L) and grown at 28°C. Two days later, single colonies were picked and streaked onto the same medium, then spread to give single colonies. Single colonies were picked again and streaked, then spread for single colonies. Single colonies were picked a third time, grown as streaks, then subjected to a quality analysis involving growth on lactose medium and chromogenic assay with Benedict's reagent. Of ten strains developed in this way, the fastest coloring colony was chosen for further work.

#### D. Transformation Of Tobacco With *Agrobacterium* Carrying Plasmid pDAB2041

Tobacco transformation with *Agrobacterium tumefaciens* was carried out by a method similar, but not identical, to published methods (R Horsch et al, 1988. Plant Molecular Biology Manual, S. Gelvin et al, eds., Kluwer Academic Publishers, Boston). To provide source tissue for the transformation, tobacco seed (*Nicotiana tabacum* cv. Kentucky 160) were surface sterilized and planted on the surface of TOB-, which is a hormone-free Murashige and Skoog medium (T. Murashige and F. Skoog, 1962). A revised medium for rapid growth and bioassays with tobacco tissue culture. Plant Physiol. 75: 473-497) solidified with agar. Plants were grown for 6-8 weeks in a lighted incubator room at 28-30°C and leaves were collected sterilely for use in the transformation protocol. Approximately one cm<sup>2</sup> pieces were sterilely cut from these leaves, excluding the midrib. Cultures of the

*Agrobacterium* strains (EHA101S containing pDAB2041), which had been grown overnight on a rotor at 28°C, were pelleted in a centrifuge and resuspended in sterile Murashige & Skoog salts, adjusted to a final optical density of 0.7 at 600 nm. Leaf pieces were dipped in this bacterial suspension for approximately 30 seconds, then blotted dry on sterile paper towels and placed right side up on medium TOB+ (Murashige and Skoog medium containing 1 mg/L indole acetic acid and 2.5 mg/L benzyladenine) and incubated in the dark at 28°C. Two days later the leaf pieces were moved to medium TOB+ containing 250 mg/L cefotaxime (Agri-Bio, North Miami, Florida) and 100 mg/L kanamycin sulfate (AgriBio) and incubated at 28-30°C in the light. Leaf pieces were moved to fresh TOB+ with cefotaxime and kanamycin twice per week for the first two weeks and once per week thereafter. Leaf pieces which showed regrowth of the *Agrobacterium* strain were moved to medium TOB+ with cefotaxime and kanamycin, plus 100 mg/l carbenicillin (Sigma). Four to six weeks after the leaf pieces were treated with the bacteria, small plants arising from transformed foci were removed from this tissue preparation and planted into medium TOB- containing 250 mg/L cefotaxime and 100 mg/L kanamycin in Magenta GA7 boxes (Magenta Corp., Chicago). These plantlets were grown in a lighted incubator room. After 3-4 weeks the primary transgenic plants had rooted and grown to a size sufficient that leaf samples could be analyzed for expression of protein from the transgene. Twenty-five independent transgenic events were recovered as single plants from the pDAB2041 transformation.

Eight independent lines expressing various levels of transgenic protein from the T-DNA of pDAB2041 were propagated *in vitro* from leaf pieces as follows. Twelve to sixteen approximately one cm<sup>2</sup> pieces were sterilely cut from leaves of each primary transgenic plant, excluding

the midrib and all naturally occurring edges. These leaf pieces were placed on medium TOB+ containing 250 mg/L cefotaxime and 100 mg/L kanamycin, and cultured in the lighted incubator at 28-30°C for 3-4 weeks, at which time small plants could be cut from the proliferating tissue mass. Several small plantlets from each transgenic line were moved into Magenta boxes containing medium TOB- plus cefotaxime and kanamycin and allowed to root and grow. The proliferating tissue mass was further cultured on medium TOB+ with cefotaxime and kanamycin, and additional plants could be cut out and grown up as needed.

Plants were moved into the greenhouse by washing the agar from the roots, transplanting into soil in 5 1/2" square pots, placing the pot into a Ziploc bag (DowBrands), placing plain water into the bottom of the bag, and placing in indirect light in a 30°C greenhouse for one week. After one week the bag could be opened; the plants were fertilized and allowed to grow further, until the plants were acclimated and the bag was removed. Plants were grown under ordinary warm greenhouse conditions (30°C, 16 H light). Plants were suitable for sampling four weeks post transplant.

### Example 3

#### 25 Characterization Of Transgenic Tobacco Plants Expressing Photorhabdus Toxin That Confer Insect Control.

##### A. Polyclonal Antibody Production

The *E. coli* produced recombinant TcdA protein was purified by a series of column purification. The protein was sent to Berkley Antibody Company (Richmond, CA) for the production of antiserum in a rabbit. Inoculations with the antigen were initiated with 0.5 mg of protein followed by four boosting injections of 0.25 mg each at about three week intervals. The rabbit serum was tested by the standard Western analysis using the recombinant TcdA protein as the antigen and enhanced chemi-

luminescens, ECL method (Amersham, Arlington Heights, IL)  
) .The antibodies (PAb-EA<sub>0</sub>) were purified using a PURE I  
antibody purification kit (Sigma, St. Luis, MO). PAb-EA<sub>0</sub>  
antibodies recognize the full-length TcdA and its  
5 processed components.

#### B. Expression Of TcdA Protein In Tobacco

Protein was extracted from the leaf tissue of  
transformed and non-transformed tobacco plants following  
the procedure described immediately below.

10 Two leaf disks of 1.4 cm in diameter were harvested  
from the middle portion of a fully expanded leaf. The  
disks were placed on a 1.6 x 4 cm piece of 3M Whatman  
paper. The paper was folded lengthwise and inserted in a  
flexible straw. Four hundred micro liters of the  
15 extraction buffer (9.5 ml of 0.2 M NaH<sub>2</sub>PO<sub>4</sub>, 15.5 ml of 0.2  
M Na<sub>2</sub>HPO<sub>4</sub>, 2 ml of 0.5 M Na<sub>2</sub>EDTA, 100 ml of Triton X100, 1  
ml of 10% Sarkosyl, 78 ml of beta-mercaptoethanol, H<sub>2</sub>O to  
bring total volume to 100 ml) was pipetted on to the  
paper. The straw containing the sample was then passed  
20 through a rolling device used for squeezing out the  
extract 1.5 mL micro centrifuge tube was placed at the  
other end of the straw to collect the extract. The  
extract was centrifuged for 10 minutes at 14,000 rpm in  
an Eppendorf refrigerated microcentrifuge. The  
25 supernatant was transferred into a new tube. Protein  
quantitation analysis was performed using the standard  
Bio-Rad Protein Analysis protocol (Bio-Rad Laboratories,  
Hercules, CA). The extract was diluted to 2 mg/ml of  
total protein using the extraction buffer.

30 For the detection of transgenic protein, Western  
blot analysis was performed. Following a standard  
procedure for protein separation (Laemmli, 1970), 40 µg  
of protein was loaded in each well of 4-20% gradient  
polyacrylamide gel (Owl Scientific Co., MA) for  
35 electrophoresis. Subsequently, the protein was

transferred onto a nitrocellulose membrane using a semi-dry electroblotter (Pharmacia LKB Biotechnology, Piscataway, NJ). The membrane was incubated for one hour in Blotto (5% milk in TBST solution; 25 mM Tris HCL pH 7.4, 136 mM NaCl, 2.7 mM KCl, 0.1% Tween 20). Thereafter, Blotto was replaced by the primary antibody solution (in Blotto). After one hour in the primary antibody, the membrane was washed with TBST for five minutes three times. Then the secondary antibody in Blotto (1:2000 dilution of goat anti-rabbit IgG conjugated to horseradish peroxidase; Bio-Rad Laboratories). was added to the membrane. After one hour of incubation, the membrane was washed with an excess amount of TBST for 10 minutes four times. The protein was visualized by using the enhanced chemi-luminescens, ECL method (Amersham, Arlington Heights, IL ). The differential intensity of the protein bands were measured using densitometer (Molecular Dynamics Inc., Sunnyvale, CA).

To determine the expression of TcdA protein in tobacco transformed with pDAB2041, PAb-EA<sub>0</sub> antibodies were used as the primary antibodies. The expression levels of TcdA protein varied among independent transformation events. The primary plant generated from the event #2041-13 showed the highest level of pre-pro TcdA expression of extractable protein. When the leaf pieces from this plant (#2041-13) were used in *in vitro* propagation, several plants were obtained. Seven of these plants were analyzed for the expression of the TcdA protein. All but one plant produced the full-length TcdA protein as well as some processed peptide components. Using the antibodies specific to Neomycin phosphotransferase, NPT (5 prime-3 prime, Boulder, Co), the expression the selectable marker gene (*npt II*) was detected. Similar results were obtained for #2041-29.

35

Table 5

-25-

Western analysis of plants derived from event #2041-13.

Plant #	TcdA	NPT (selectable marker)
2041-13A	+	not done
2041-13B	+	not done
2041-13-1	-	+
2041-13-2	+	+
2041-13-3	+	+
2041-13-4	+	+
2041-13-5	+	+

### C. Nucleic Acid Analysis of Transgenic Tobacco Lines

Genomic DNA was prepared from a group of 2041  
 5 transgenic events. The lines included Magenta box stage  
 2041-13, and greenhouse stage plants 2041-13-1, 2041-13-  
 2, 2041-13-5, 2041-9, 2041-20A and 2041-20B. A  
 transgenic GUS line (2023) was included as a negative  
 control. Southern analysis of these lines was performed.  
 10 The genomic tobacco DNA was restricted with the enzyme  
 SstI which should result in a 8.9 kb hybridization  
 product when hybridized to a *tcdA* gene specific probe.  
 The 8.9 kb hybridization product should consist of the  
 35T promoter and the *tcdA* coding region. All 2041 plants  
 15 contained a band of the expected size. Events 2041-9 and  
 -20 appear to be the same line with 5 identical  
 hybridizing bands. Event 2041-13 produced 6  
 hybridization fragments with the *tcdA* coding region  
 probe. Magenta box and various greenhouse plants of  
 20 2041-13 all produced the same hybridization profile.  
 This hybridization pattern was different from that of  
 events 2041-9 and -20.

RNA analysis, using the *tcdA* coding region probe,  
 was performed on the same group of greenhouse 2041  
 25 plants. Immunoblot analysis had revealed that plants  
 2041-9, 2041-20A, 2041-20B, and 2041-13-1 produced no  
 detectable TcdA protein; while 2041-13-2 and 2041-13-5  
 produced substantial amounts of full-length TcdA.

Northern analysis was in agreement with the immunoblot

result. A faint RNA signal was detected for plants 2041-9, 2041-20A, 2041-20B, and 2041-13-1. Only faintly visible was a band corresponding to full-length *tcdA* transcript in plant 2041-13.1. In contrast, for plants 5 2041-13-2 and 2041-13-5 a strong RNA signal was detected, with a substantial amount of full-length size (~8.0 kb) *tcdA* transcript. These data support the observed bioassay activity for this group of plants.

Genomic DNA was prepared from a second functionally 10 active 2041 transgenic event, 2041-29. Southern analysis of this line was performed. A transgenic GUS line (2023) was included as a negative control, DNA of line 2041-9 was included as a positive control.

The genomic tobacco DNAs were restricted with the 15 enzyme *Sst*I which should result in a 8.9 kb hybridization product when hybridized to a *tcdA* gene specific probe. The 8.9 kb hybridization product should consist of the 35T promoter and the *tcdA* coding region. For plant 2041-29-5, three hybridization products larger than 8.9 kb the 20 were detected with the *tcdA* gene specific probe. Immunoblot analysis has demonstrated pre-pro TcdA protein is made by this plant, it is therefore likely that a restriction site was lost during transformation or regeneration, or the 2041-29 genomic DNA was not 25 thoroughly digested.

#### D. Tobacco Leaf-Disk Tests With Tobacco Hornworm Exhibiting Insect Control

Leaves were sampled from tobacco plants, *Nicotiana* 30 *tabaco*, previously transplanted into the greenhouse. A single leaf was sampled from each plant on each test date. Leaves were selected from the zone where younger elongate leaves transition into older ovate leaves. Excised leaves were placed into 12 oz. cups with the 35 petiole submerged in water to maintain turgor, and transported to the laboratory.



Eight, 1.4 cm disks were cut from the center portion of one side of each leaf (right adaxial side up, with distal portion facing away from the observer). Each disk was placed individually into a well of a C-D

5 International 128 well tray (Pitman, NJ.) into which 0.5 ml of a 1.6% aqueous agar solution had been previously pipetted. The solidified agar prevented the leaf disks from drying out. The adaxial surface of the disk was always oriented up.

10 A single neonate tobacco hornworm, *Manduca sexta*, was placed on each disk and the wells were sealed with vented plastic lids. The assay was held at 27°C and 40% RH. Larval mortality and live-weight data were collected after 3 days. Data were subjected to analysis of

15 variance and Duncan's multiple range test ( $\alpha = 0.05$ ) (Proc GLM, SAS Institute Inc., Cary, NC.). Data were transformed using a logarithmic function to correct a correlation between the magnitude of the mean and variance.

20

Table 6  
Results of leaf-disk assays from greenhouse grown tobacco plants with event 2041-13.

TRT	Plant	Plant Age	Weight of Surviving Larvae (mg) & Duncan's Group <sup>1</sup>				
			Pretes t	Test 1	Test 2	Test 3	3 Test Sum.
13	non-transformed - 2	young	---	---	---	18.8 a*	---
14	non-transformed - 3	young	---	---	---	17.0 ab	---
16	non-transformed - 5	young	---	---	---	16.4 ab	---
3	2041-13-1 (western -)	young	---	17.6 a	18.2 a	16.1 ab	17.3 a
9	Gus Control	old	19.3 a	14.6 a	16.3 a	14.5 ab	15.1 a
10	non-transformed - 1	young	---	8.3 b	16.8 a	13.9 b	13.0 b
11	2041-20B (western -)	old	---	10.0 b*	13.7 ab	14.6 ab	12.9 b
15	non-transformed - 4	young	---	---	---	13.0 bc	---
8	2041-20A (western -)	old	15.7 a	8.3 b	11.3 bc	9.2 cd	9.6 c
12	2041-9 (western -)	old	19.5 a	---	---	7.9 d	---
7	2041-13-5 (western +)	young	---	6.3 bc	9.6 cd	7.2 de	7.7 d
5	2041-13-3 (western +)	young	---	6.4 bc****	6.2 e	6.8 de**	6.4 de
1	2041-13A (western +)	old	7.2 b	6.8 bc*	7.0 de*	5.4 e	6.4 de
6	2041-13-4 (western +)	young	---	4.9 c****	5.8 e	7.6 d	6.4 de
4	2041-13-2 (western +)	young	---	5.7 bc	5.7 e**	7.5 d	6.3 de
2	2041-13B (western +)	old	---	4.7 c**	5.6 e	7.2 de	5.9 e

\* Number of stars corresponds to the number of dead larvae per 8 tested.

1. Data transformed (logarithm) for analysis.  
Means followed by the same letter are not significantly different ( $\alpha = 0.05$ ).

5  
TABLE 7  
Results Of Leaf-Disk Assays From Greenhouse Grown Tobacco Plants  
With Event 2041-29.

Plant	MEAN WGT (MG) / Duncan's Group				Four Test Summary
	Test 1	Test 2	Test 3	Test 4	
2014-6 GUS 1	15.8 a	16.6a	**5.5bc	*12.9ab	13.2 a
2014-6 GUS 2	14.4 a	*6.6 bc	*13.4a	15.2a	12.6 a
KY-160 NTC	13.4 a	6.7 bc	7.9b	8.5bc	9.1 b
2041-29 4P	*4.9 b	*7.3b	****6.9b	*****	6.3 c
2041-29 7	*5.9 b	5.1bc	***6.7b	***7.2c	6.1 c
2041-29 3P	*5.6 b	**7.9b	*****6.5b	***3.6d	5.9 c
2041-29 2P	6.3 b	****4.7c	*****4.1c	*****4.6d	5.4 c

10 \* Number of stars corresponds to the number of dead larvae per 8 tested.

1. Data transformed (logarithm) for analysis.

Means followed by the same letter are not significantly different ( $\alpha = 0.05$ ).

15 All event 2041-29 plants significantly depressed THW larval weight gain compared to control plants. Average weight depression was 49%. Statistically significant mortality occurred in THW larvae exposed to foliage from 2041-29 plants. Mortality averaged 37.5% compared to 5.2% in controls.

20

#### E. Isolation and Characterization of Functional Photorhabdus Toxin Protein From Transgenic Plants

25 Seven grams of transgenic tobacco plants (2041-13) expressing TcdA (Toxin A) gene were homogenized with 10 ml 50 mM Potassium Phosphate buffer, pH 7.0 using a bead beater (Biospec Products, Bartlesville, OK) according to manufacturer's instructions. The homogenate was filtered through four layers of cheese cloth and then centrifuged at 35,000 g for 15 min. The supernant was collected and 30 filtered through 0.22  $\mu$ m Millipore Express™ membrane. It was then applied to a Superdex 200 cloumn (2.6 x 40 cm)

which had been equilibrated with 20 mM Tris buffer, pH 8.0 (Buffer A). The protein was eluted in Buffer A at a flow rate of 3 ml/min. Fractions with 3 ml each were collected and subjected to southern corn rootworm (SCR) bioassay. It was found that fractions corresponding to a native molecular weight around 860 kDa had the highest insecticidal activity. Western analysis of the active fraction using a polyclonal antibody specific to Toxin A indicated the presence of full-length TcdA peptide. The active fractions were further combined and applied to a Mono Q 10/10 column which had been equilibrated with Buffer A. Proteins bound to the column were then eluted by a linear gradient of 0 to 1 M NaCl in Buffer A. Fractions with 2 ml each were collected and analyzed by both SCR bioassay and Western using antibody specific to Toxin A. The results again demonstrated the correlation between insecticidal activity and presence of full-length TcdA peptide.

## 20 F. Characterization of Progeny Transgenic Plants

The inheritability of the genetically engineering plants containing the *Photorhabdus* toxin gene was evaluated by generating F1 progeny. Progeny was generated from 2041-13 event by selfing expression positive plants. The 2041-13 plants in the greenhouse were allowed to self-pollinate. Seed capsules were collected when mature and were allowed to dry and after-ripen on the laboratory bench for two weeks. Seed from plant designated 2041-13A was surface-sterilized and distributed on the surface of medium TOB- without selection, to allow recovery of nonexpressing or nontransgenic progeny as well as expressing and segregating transgenic siblings. Seed was germinated in a C lighted incubator room (16 H light, 28 C). After 1 month, fifty-one seedlings, designated 2041-13A-S1 through S51, were distributed into Magenta boxes

self-fertilized 2041-13 plants genetically engineered to produce the "204" A toxin. The tests included 6 non-expressing progeny (protein-negative controls), 45 toxin A expressors, and 4 non-transformed controls (KY-160).

- 5 Results are from three leaf-disk assays (method previously outlined) where eight disks were used per test. The data were analyzed using analysis of variance and were blocked by test.

The treatment effect for each of these analyses indicated the  $Pr > F$  was less than 0.0001. The Toxin A expressors produced significant control of tobacco hornworm compared to each of the control groups based on each of the three measures of efficacy. The two control groups behaved similarly. Statistical analysis using ANOVA and an LSD test with alpha equal to 0.01 (or 1%) showed differences between the 3 groups. The LSD test indicated that the non-expressors and the non-transformed plants were similar in larvae weights but the expressors gave weights significantly lower than either of the other two groups of plants. These data demonstrated that the genetic basis for insect control was inheritable and corresponded to the presence of expressed toxin gene.

25 Table 8  
Tobacco hornworm results from F1 progeny of self-fertilized  
2041-13 tobacco plants.

Treatment Group	Mean Value and Duncan's Grouping <sup>d</sup>		
	Total Weight (mg) <sup>a</sup>	Survivor Weight (mg) <sup>b</sup>	Leaf Area (cm <sup>2</sup> ) <sup>c</sup>
Non-transformed Control	15.8 a	15.8 a	1.2 a
Protein-negative Control	16.4 a	16.5 a	1.2 a
Toxin A Expressor	8.1 b	9.2 b	4.9 b

<sup>a</sup> Average insect weight with dead insects considered to weigh nothing.

<sup>b</sup> Average insect weight with dead insects excluded from analysis.

<sup>c</sup> Total leaf area remaining per eight leaf disks. Initial area was approximately 12 cm<sup>2</sup>.

<sup>d</sup> Means followed by the same letter are not significantly different (alpha = 0.05).

## Example 4

5 Transformation Of Maize With a Vector Carrying Plasmid  
pDAB1834 Encoding *Photorhabdus* Toxins

A. Preparation Of Maize Transformation Vectors  
Containing Modified Plant-Optimized *TcdA* Coding Regions:  
Plasmid Pdab1834

10

Preparation of maize transformation vectors was accomplished in two steps. First, a modified plant-optimized *tcdA* coding region was ligated into a plant expression cassette plasmid. In this step, the coding  
15 region was placed under the transcriptional control of a promoter functional in maize plant cells. RNA transcription termination and polyadenylation were mediated by a downstream copy of the terminator region from the *Agrobacterium* nopaline synthase gene. One  
20 plasmid designed to function in this role is pDAB1538. In the second step, the complete gene comprised of the promoter, coding region, and 3' UTR terminator region was ligated to a plant transformation vector that contained a plant expressible selectable marker gene which allowed  
25 the selection of transformed maize plant cells amongst a background of nontransformed cells. An example of such a vector is pDAB367.

It is a feature of plasmid pDAB1538 that any coding region having an *NcoI* site at its 5' end and a *SacI* site  
30 3' to the coding region, when cloned into the unique *NcoI* and *SacI* sites of pDAB1538, is placed under the transcriptional control of the maize ubiquitin1 (*ub1*) promoter. It is also a feature of pDAB1538 that the 5' untranslated leader (UTR) sequence preceding the *NcoI*  
35 site comprises a polylinker. Additionally it is a feature of pDAB1538 that transcription termination and polyadenylation of the mRNA containing the introduced coding region are mediated by termination/Poly A addition

sequences derived from the nopaline synthase (Nos) gene. Finally; it is a feature of pDAB1538 that the entire assembly of promoter/coding region/3'UTR can be obtained as a single DNA fragment by cleavage at the flanking *NotI* sites.

It is a feature of pDAB367 that the phosphinothricin acetyl transferase protein, which has as its substrate phosphinothricin and related compounds, is produced in plant cells through transcription of its coding region mediated by the Cauliflower Mosaic Virus 35S promoter and that termination of transcription plus polyadenylation are mediated by the nopaline synthase terminator region. It is further a feature of pDAB367 that any DNA fragment containing flanking *NotI* sites can be cloned into the unique *NotI* site of pDAB367, thus physically linking the introduced DNA fragment to the aforementioned selectable marker gene.

To prepare a maize plant-expressible gene to produce the endoplasmic reticulum-targeted TcdA protein in plant cells, DNA of a plasmid (pA0H\_4-ER) containing the plant-optimized, ER-targeted *tcdA* coding region, (SEQ ID No:6) was cleaved with restriction enzymes *NcoI* and *SacI*, and the large 7610 bp fragment was ligated to similarly-cut DNA of plasmid pDAB1538 to produce plasmid pDAB1832. DNA of pDAB1832 was then digested with *NotI*, and the 9984 bp *NotI* fragment was ligated into the unique *NotI* site of pDAB367 to produce plasmid pDAB1834.

It is a feature of plasmids pDAB1834 that the *ubil* and 35S promoters are encoded on the same DNA strand.

#### B. Transformation and Regeneration of Transgenic Maize Isolates

Type II callus cultures were initiated from immature zygotic embryos of the genotype "Hi-II." (Armstrong et al, (1991) Maize Genet. Coop. Newslett., 65: 92-93). Embryos were isolated from greenhouse-grown ears from

crosses between Hi-II parent A and Hi-II parent B or F<sub>2</sub> embryos derived from a self- or sib-pollination of a Hi-II plant. Immature embryos (1.5 to 3.5 mm) were cultured on initiation medium consisting of N6 salts and vitamins (Chu et al, (1978) *The N6 medium and its application to anther culture of cereal crops*. Proc. Symp. Plant Tissue Culture, Peking Press, 43-56), 1.0 mg/L 2,4-D, 25mM L-proline, 100 mg/L casein hydrolysate, 10 mg/L AgNO<sub>3</sub>, 2.5 g/L GELRITE (Schweizerhall, South Plainfield, NJ), and 20 g/L sucrose, with a pH of 5.8. After four to six weeks callus was subcultured onto maintenance medium (initiation medium in which AgNO<sub>3</sub> was omitted and L-proline was reduced to 6 mM). Selection for Type II callus took place for ca. 12-16 weeks.

Plasmid pDAB1834 was transformed into embryogenic callus. For blasting, 140 µg of plasmid DNA was precipitated onto 60 mg of alcohol-rinsed, spherical gold particles (1.5 - 3.0 µm diameter, Aldrich Chemical Co., Inc., Milwaukee, WI) by adding 74 µL of 2.5M CaCl<sub>2</sub> H<sub>2</sub>O and 30 µL of 0.1M spermidine (free base) to 300 µL of plasmid DNA and H<sub>2</sub>O. The solution was immediately vortexed and the DNA-coated gold particles were allowed to settle. The resulting clear supernatant was removed and the gold particles were resuspended in 1 ml of absolute ethanol. This suspension was diluted with absolute ethanol to obtain 15 mg DNA-coated gold/mL.

Approximately 600 mg of embryogenic callus tissue was spread over the surface of Type II callus maintenance medium as described herein lacking casein hydrolysate and L-proline, but supplemented with 0.2 M sorbitol and 0.2 M mannitol as an osmoticum. Following a 4 h pre-treatment, tissue was transferred to culture dishes containing blasting medium (osmotic media solidified with 20 g/L TC agar (PhytoTechnology Laboratories, LLC, Shawnee Mission, KS) instead of 7 g/L GELRITE. Helium blasting accelerated suspended DNA-coated gold particles towards

and into the prepared tissue targets. The device used was an earlier prototype of that described in US Patent 5,141,131 which is incorporated herein by reference. Tissues were covered with a stainless steel screen (104  
5  $\mu\text{m}$  openings) and placed under a partial vacuum of 25 inches of Hg in the device chamber. The DNA-coated gold particles were further diluted 1:1 with absolute ethanol prior to blasting and were accelerated at the callus targets four times using a helium pressure of 1500 psi,  
10 with each blast delivering 20  $\mu\text{L}$  of the DNA/gold suspension. Immediately post-blasting, the tissue was transferred to osmotic media for a 16-24 h recovery period. Afterwards, the tissue was divided into small pieces and transferred to selection medium (maintenance  
15 medium lacking casein hydrolysate and L-proline but containing 30 mg/L BASTA® (AgrEvo, Berlin, Germany)). Every four weeks for 3 months, tissue pieces were non-selectively transferred to fresh selection medium. After 7 weeks and up to 22 weeks, callus sectors found  
20 proliferating against a background of growth-inhibited tissue were removed and isolated. The resulting BASTA®-resistant tissue was subcultured biweekly onto fresh selection medium. Following western analysis, positive transgenic lines were identified and transferred to  
25 regeneration media. Western-negative lines underwent subsequent RNA spot blot analysis to identify negative controls for regeneration.

Regeneration was initiated by transferring callus tissue to cytokinin-based induction medium, which  
30 consisted of Murashige and Skoog salts, hereinafter MS salts, and vitamins (Murashige and Skoog, (1962) *Physiol. Plant.* 15: 473-497) 30 g/L sucrose, 100 mg/L myo-inositol, 30 g/L mannitol, 5 mg/L 6-benzylaminopurine, hereinafter BAP, 0.025 mg/L 2,4-D, 30 mg/L BASTA®, and  
35 2.5 g/L GELRITE at pH 5.7. The cultures were placed in low light (125 ft-candles) for one week followed by one



week in high light (325 ft-candles). Following a two week induction period, tissue was non-selectively transferred to hormone-free regeneration medium, which was identical to the induction medium except that it lacked 2,4-D and BAP, and was kept in high light. Small (1.5-3 cm) plantlets were removed and placed in 150x25 mm culture tubes containing SH medium (SH salts and vitamins (Schenk and Hildebrandt, (1972) Can. J. Bot. 50:199-204), 10 g/L sucrose, 100 mg/L myo-inositol, 5 mL/L FeEDTA, and 2.5 g/L GELRITE, pH 5.8). Plantlets were transferred to 12 cm pots containing approximately 0.25 kg of METRO-MIX 360 (The Scotts Co. Marysville, OH) in the greenhouse as soon as they exhibited growth and developed a sufficient root system. They were grown with a 16 h photoperiod supplemented by a combination of high pressure sodium and metal halide lamps, and were watered as needed with a combination of three independent Peters Excel fertilizer formulations (Grace-Sierra Horticultural Products Company, Milpitas, CA). At the 6-8 leaf stage, plants were transplanted to five gallon pots containing approximately 4 kg METRO-MIX 360, and grown to maturity.

#### EXAMPLE 5

##### Characterization Of Transgenic Maize Plants

Expressing Photorhabdus Toxin That Confer Insect Control.  
A. Insect Bioassays

A single leaf was sampled from each plant in each test. Eight, 1.4 cm disks were cut from the outer portion of each leaf (approximately 30cm long) avoiding the center vein. Each disk was placed individually into a well of a C-D International 128 well tray (Pitman, NJ.) into which 0.5 ml of a 1.6% aqueous agar solution had been previously pipetted. The solidified agar prevented the leaf disks from drying out. The adaxial surface of the disk was always oriented up.

Five neonate southern corn rootworms, *Diabrotica undecimpunctata howardi*, were placed on each disk and the wells were sealed with vented plastic lids. The assay was held at 27°C and 40% RH. Larval mortality and live-weight data were collected after 3 days. Data were subjected to analysis of variance and Duncan's multiple range test ( $\alpha = 0.05$ ) (Proc GLM, SAS Institute Inc., Cary, NC.). Weight data were transformed using a logarithmic function to correct a correlation between the magnitude of the mean and variance.

TABLE 9

Results of Maize Leaf-disk Test vs SCR

Treatment	Mean % Kill (Duncan's)	Mean Survival Weight (mg) (Duncan's)
1834 - 11	68 A	0.064 A
1834 - 17	44 B	0.098 B
1834 - 15	26 BC	0.127 C
HiII control	13 C	0.161 C

Note: Means followed by the same letter are not significantly different based on Duncan's multiple range test ( $\alpha=0.05$ ). Insect groups weighing less than 0.1 mg were set to 0.03 mg instead of zero to conduct a more conservative analysis. Mortality ( $\arcsin(\sqrt{\text{mortality}})$ ) and weight ( $\log_{10}$ ) data were transformed for analyses.

20

The results shown in Table 9 demonstrated that two events expressing TcdA protein were statistically distinct from control lines bioassayed using SCR neonates by mortality and survival weight criteria. These results demonstrated that southern corn rootworm were functionally effected by feeding on maize plants containing and expressing the *tcdA* gene. Those plants from 1834-11 were used to generate progeny for testing of inheritability of transgene.

B. PRODUCTION AND PROGENY TEST OF *tcdA* TRANSGENIC MAIZE

Origin and growth of progeny plants: Sibling plants 1834-11-07 and 1834-11-08, clonally derived by regeneration from the callus of transgenic maize event 1834-11, were transplanted to the greenhouse and pollinated with inbred OQ414. Seeds obtained from these crosses, comprising seed lots 1834-11-07A and 1834-11-08A, were planted in Roottrainers (1 ½ inch x 2 inch x 8 inch deep, product #647, C. Hummert Intl., Earth City, Mo.) filled with Metro-Mix 360 soilless mix (Scotts Terra-Lite, available from Hummert Intl.) and top irrigated with Hoagland's nutrient solution. (Hoagland's solution contains 229 ppm nitrogen as nitrate, 24.6 ppm nitrogen as ammonium, 26 ppm P, 157 ppm K, 187 ppm Ca, 49 ppm Mg. and 30 ppm Na.)

Greenhouse conditions for this trial were: 16 hour days, daylight supplemented by metal halide lamps as needed to achieve a minimum of 600 ?Einsteins/cm<sup>2</sup> PAR, and ambient temperature 30 C days, 22 C nights.

Leaves were sampled for protein determination approximately one week after planting. Leaf bioassays were conducted 2-3 weeks after planting; root bioassays were initiated approximately 3 weeks post planting.

Protein analysis of progeny plants: Protein was extracted from leaf and root samples harvested from transgenic plants, line 1834-11 progenies, and non-transformed plants. Each sample was placed on a 1.6 x 4 cm piece of 3M Whatman<sup>TM</sup> paper. The paper was folded lengthwise and inserted in a flexible straw. A volume of 350 µl of an extraction buffer (9.5 ml of 0.2 M NaH<sub>2</sub>PO<sub>4</sub>, 15.5 ml of 0.2 M Na<sub>2</sub>HPO<sub>4</sub>, 2 ml of 0.5 M Na<sub>2</sub>EDTA, 100 ml of Triton X-100, 1 ml of 10% Sarkosyl, 78 ml of beta-mercaptoethanol, H<sub>2</sub>O to bring total volume to 100 ml, 50 µg/ml Antipain, 50 µg/ml Leupeptin, 0.1 mM Chymostatin, 5 µg/ml Pepstatin) was pipetted on to the paper. The straw containing the

sample was then passed through a rolling device used for squeezing the extract into a 1.5 ml microcentrifuge tube. The extract was centrifuged for 10 minutes at 14,000 rpm in an Eppendorf refrigerated micro-centrifuge. The  
5 supernatant was transferred into a new tube. The amount of the total extractable protein was determined using a standard BioRad Protein Analysis protocol (BioRad Laboratories, Hercules, CA).

The presence of the TcdA protein was visualized by  
10 Western blot analysis following a standard procedure for protein separation (Laemmli, 1970). A volume of twenty  $\mu$ l of extract was loaded in each well of 4-20% gradient polyacrylamide gel (Owl Scientific Co., MA) for electrophoresis. Subsequently, the protein was  
15 transferred onto a nitrocellulose membrane using a semi-dry electroblotter (Pharmacia LKB Biotechnology, Piscataway, NJ). The membrane was incubated for one hour in TBST-M solution (10% milk in TBST solution; 25 mM Tris HCL pH 7.4, 136 mM NaCl, 2.7 mM KCl, 0.1% Tween 20).  
20 Thereafter, the primary antibody (Anti-TcdA in TBST-M) was added. After one hour, the membrane was washed with TBST for five minutes, three times. Then the secondary antibody solution (goat anti-rabbit IgG conjugated to horseradish peroxidase; Bio-Rad Laboratories, in TBST-M)  
25 was added to the membrane. After one hour of incubation, the membrane was washed with an excess amount of TBST for 10 minutes, four times. The protein was visualized using the Super Signal® West Pico chemiluminescence method (Pierce Chemical Co., Rockford, IL). The protein blot  
30 was exposed on a Hyper-film (Amersham, Arlington Heights, IL) and was developed within 3 minutes. The intensity of the protein band was measured using a densitometer (Molecular Dynamics Inc., Sunnyvale, CA) and compared to standards.

35 Three of six plants from seed lot 1834-11-07A and three of six plants from seed lot 1834-11-08A produced

detectable levels of TcdA protein (Table 1).

Approximately 3.8 to 13.3 ppm of TcdA were detected in the leaf blades and 4.1 to 8.4 ppm were detected in the leaf tips of the protein-positive plants. The amounts of  
5 TcdA protein detected in the roots were slightly lower than those found in the leaves.

Insect bioassays with progeny plants: Plants were selected for bioassay based on results from Western blot  
10 analysis. Twelve (12), 6.4 mm diameter leaf discs were cut from the youngest leaf of each 2 week old seedling. Each disc was placed in a well of a 128-well tray (CD International) containing approximately 0.5mL of a solidified 2% agar in water solution. Two neonate  
15 southern corn rootworm, *Diabrotica undecimpunctata howardi* (Barber) (SCR), were placed in each well with a leaf disc. Trays were covered with perforated lids and maintained under a controlled environment for 3 days (28 C; 16 hours light:8 hours dark; approx. 60% relative  
20 humidity). Living larvae from 4 leaf discs were pooled and weighed producing 3 weight determinations per plant. Average weights were calculated by dividing the pooled weight by the number of survivors. Differences in average weights of SCR fed leaf discs from protein  
25 positive and protein negative plants were assessed using analysis of variance on the natural log-transformed average weights (Minitab, v. 12.2, Minitab Inc., State College, PA).

30 Root bioassays were initiated approximately 1 week after the initiation of the leaf disc bioassays. Approximately 24h prior to eclosion, SCR eggs were suspended in a 0.15% solution of agar in water to a concentration of 100 eggs/ml. Plants were inoculated  
35 with SCR eggs by pipetting 2.0 ml of the egg suspension (ie., approximately 200 eggs) just below the soil surface at the base of each plant. Two weeks after inoculation, plants were removed from their Roottrainer pots, their

roots washed free of potting mix, and scored for rootworm damage based on a 1 (resistant) to 9 (susceptible) rating system (Welch, 1977). The results of the root ratings were examined using non-parametric tests to determine if the distribution of root ratings from the protein positive plants was the same as the distribution of the ratings from the protein negative plants. Testing was done at the 5% significance level. (StatXact v.3, CYTEL Software Corporation, Cambridge MA)

Results from leaf and root bioassays of tcdA protein positive and protein negative progeny plants are summarized in Table 10. The average weights of SCR larvae fed leaf discs from protein positive plants were significantly lower than those of larvae fed leaf discs from protein negative plants ( $F = 4.6$ ; d.f. = 1, 34;  $P \leq 0.001$ ). The Kolmogorov-Smirnov 2 sample test ( $p=0.04$ ) and the Wald Wolfowitz runs test ( $p=0.001$ ) indicated that the protein positive and protein negative root rating distributions were not similar. The Wilcoxon-Mann-Whitney test ( $p=0.0206$ ) and the Normal Scores test ( $p=0.206$ ) indicated that the average score for the protein positive plants was lower than the average root rating from the protein negative plants.

Table 10. Protein analysis and insect bioassay results with progeny of TcdA transgenic maize.

Plant Number	TcdA Protein	Leaf Disc Bioassay Avg. Wt. (mg)	Root Bioassay Root Rating (1-9)
1834-11-07A-30	PRO-	0.190	8
1834-11-08A-21	PRO-	0.196	9
1834-11-08A-16	PRO-	0.195	9
1834-11-08A-14	PRO-	0.137	9
1834-11-07A-22	PRO-	0.208	9
1834-11-07A-20	PRO-	0.175	9

1834-11-07A-26	PRO+	0.118	9
1834-11-08A-17	PRO+	0.132	8
1834-11-07A-14	PRO+	0.110	2
1834-11-07A-11	PRO+	0.106	4
1834-11-08A-28	PRO+	0.129	8
1834-11-08A-27	PRO+	0.108	4

DNA analysis of progeny plants: Leaf samples from 1834-11.7A and 1834-11.8A progeny plants were in conical 50 ml polypropylene tubes and dried in a Labconco Freeze Dry

5 Lyophilizer (Kansas City, MO) for 1-2 days. Lyophilized leaves were then ground in a Tecator Cyclotec 1093 Sample mill grinder (Hoganas, Sweden) and stored at -20C.

Genomic DNA was extracted by the following procedure: (1) to a 25 ml Conical tube containing 300-500 mg of ground

10 tissue, 9 ml of CTAB (cetyl trimethylammonium bromide solution) was added, and incubated at 65°C for 1 hour; (2) 4.5 ml of chloroform: octanol (24:1) was added and mixed gently for 5 minutes; (3) samples were centrifuged at 2000 rpm and DNA was precipitated from the supernatant

15 with an equal volume of isopropanol; (4) DNA was collected on a glass hook, washed in ethanol, and dissolved in TE (10 mM Tris.HCl, 0.5 mM EDTA, pH8.0).

Genomic DNA was digested at 37 °C. for 2 hours in an

20 Eppendorf tube containing the following mixture:

8 µl of 800ug/ml DNA, 2 µl 1 mg/ml BSA (Bovine serum albumin), 2 µl 10x buffer, 1 µl SacI, 1 µl EcoRI, and 6 µl H<sub>2</sub>O. Digested DNA samples were electrophoresed overnight at 40 mA in a 0.85% SeaKem LE agarose gel (FMC, Rockland,

25 Maine). The gel was blotted onto Millipore Immobilon-Ny+ (Bedford, MA) membrane overnight in 20X SSC (NaCl 175.2 g/l, Na citrate 88 g/l). The probe DNA was cut with BamHI/SacI (NEB, Beverly, MA) from pDAB1551 plasmid, which released a 7356 bp fragment containing the open

30 reading frame of the rebuilt *tcdA* gene. This 7356 bp fragment was labeled with P32 using a Stratagene Prime-it

RmT dCTP-Labeling Reactions kit (La Jolla, CA) and used for Southern hybridization. Hybridization was conducted in hybridization buffer (10% polyethylene glycol, 7% SDS [Sodium dodecyl sulfate], 0.6X SSC, 10 mM NaPO<sub>4</sub>, 5 mM EDTA, 10 µg/ml denatured salmon sperm) at 60 °C overnight. After hybridization, the membrane was washed with 10X SSC plus 0.1% SDS at 60 °C for 30 min and exposed to X ray film (Hyperfilm® MP, Amersham Life Sciences, Piscataway, NJ) for 1-2 days.

10

Results summarized indicate that a pattern of 8 hybridizing bands (the size of the expected fragment and larger) cosegregated with protein expression in 50% of all progeny assayed. These results are characteristic of a complex insertion at a single site. All seedlings containing the insert also expressed toxin protein.

15

#### Example 6

Transformation Of Rice With a Vector Carrying Plasmid pDAB1553 Encoding *Photobacterium* Toxins

20

##### A. Plasmid pDAB1553

Plasmid pDAB1553 containing *tcdA* driven by the maize ubiquitin1 promoter and *hpt* (hygromycin phosphotransferase providing resistance to the antibiotic hygromycin) under the control of 35T (a modified 35S promoter), was used for transformation.

25

Preparation of rice transformation vectors was accomplished in two steps. First, a modified plant-optimized *tcdA* coding region was ligated into a rice plant expression cassette plasmid. In this step, the coding region was placed under the transcriptional control of a promoter functional in plant cells. RNA transcription termination and polyadenylation were mediated by a downstream copy of the terminator region from the *Agrobacterium* nopaline synthase gene. One

35



plasmid designed to function in this role is plasmid pDAB1538 (described in the section on maize transformation vectors). In the second step, the complete gene comprised of the promoter, coding region, and terminator region was ligated to a rice plant transformation vector that contained a plant expressible selectable marker gene which allowed the selection of transformed rice plant cells amongst a background of nontransformed cells. An example of such a vector is pDAB354-NotI.

It is a feature of pDAB354-NotI that the hygromycin phosphotransferase protein, which has as its substrate hygromycin B and related compounds, is produced in plant cells through transcription of its coding region mediated by the Cauliflower Mosaic Virus 35S promoter and that termination of transcription plus polyadenylation are mediated by the nopaline synthase terminator region. It is further a feature of pDAB354-NotI that any DNA fragment containing flanking NotI sites can be cloned into the unique NotI site of pDAB354-NotI, thus physically linking the introduced DNA fragment to the aforementioned selectable marker gene.

To prepare a plant-expressible gene to produce the non-targeted TcdA protein in rice plant cells, DNA of a plasmid (pA0H\_4-OPTI) containing the plant-optimized *tcdA* coding region, (SEQ ID No:3) was cleaved with restriction enzymes *NcoI* and *SacI*, and the large 7550 bp fragment was ligated to similarly-cut DNA of plasmid pDAB1538 to produce plasmid pDAB1551. DNA of pDAB1551 was then digested with *NotI*, and the large 9933 bp fragment was ligated to *NotI* digested DNA of pDAB354-NotI to produce plasmid pDAB1553.

It is a feature of plasmid pDAB1553 that the *ubi1* and 35S promoters are encoded on the same DNA strand.

#### B. Production of Rice transgenics

For initiation of embryogenic callus, mature seeds of a *Japonica* cultivar, Taipei 309 were dehusked and surface-sterilized in 70% ethanol for 2-5 min. followed by a 30-45 min soak in 50% commercial bleach (2.6% sodium hypochlorite) with a few drops of 'Liquinox' soap. The seeds were then rinsed 3 times in sterile distilled water and placed on filter paper before transferring to 'callus induction' medium (i.e., NB). The NB medium consisted of N6 macro elements (Chu, 1978, The N6 medium and its application to anther culture of cereal crops. Proc. Symp. Plant Tissue Culture, Peking Press, p43-56), B5 micro elements and vitamins (Gamborg et al., 1968, Nutrient requirements of suspension cultures of soybean root cells. Exp. Cell Res. 50: 151-158), 300 mg/L casein hydrolysate, 500 mg/L L-proline, 500 mg/L L-glutamine, 30 g/L sucrose, 2 mg/L 2,4-dichloro-phenoxyacetic acid (2,4-D), and 2.5 g/L gelrite (Schweizerhall, NJ) with the pH adjusted to 5.8. The mature seed cultured on 'induction' media were incubated in the dark at 28°C. After 3 weeks of culture, the emerging primary callus induced from the scutellar region of mature embryo was transferred to fresh NB medium for further maintenance.

About 140 µg of plasmid pDAB1553 DNA was precipitated onto 60 mg of 1.0 micron (Bio-Rad) gold particles as described herein.

For helium blasting, actively growing embryogenic callus cultures, 2-4 mm in size, were subjected to a high osmoticum treatment. This treatment included placing of callus on NB medium with 0.2 M mannitol and 0.2 M sorbitol (Vain et al., 1993, Osmoticum treatment enhances particle bombardment-mediated transient and stable transformation of maize. Plant Cell Rep. 12: 84-88) for 4 h before helium blasting. Following osmoticum treatment, callus cultures were transferred to 'blasting' medium (NB+2% agar) and covered with a stainless steel screen (230 micron). The callus cultures were blasted at

2,000 psi helium pressures twice per target. After blasting, callus was transferred back to the media with high osmoticum overnight before placing on selection medium, which consisted NB medium with 30 mg/L

5 hygromycin. After 2 weeks, the cultures were transferred to fresh selection medium with a higher concentration of selection agent, i.e., NB+50mg/L hygromycin (Li et al., 1993, An improved rice transformation system using the biolistic method. Plant Cell Rep. 12: 250-255).

10 Compact, white-yellow, embryogenic callus cultures, recovered on NB+50 mg/L hygromycin, were regenerated by transferring to 'pre-regeneration' (PR) medium + 50 mg/L hygromycin. The PR medium consisted of NB medium with 2 mg/L benzyl aminopurine (BAP), 1 mg/L naphthalene acetic

15 acid (NAA), and 5 mg/L abscisic acid (ABA). After 2 weeks of culture in the dark, they were transferred to 'regeneration' (RN) medium. The composition of RN medium is NB medium with 3 mg/L BAP, and 0.5 mg/L NAA. The cultures on RN medium were incubated for 2 weeks at

20 28° C under high fluorescent light (325-ft-candles). The plantlets with 2 cm shoot were transferred to 1/2 MS medium (Murashige and Skoog, 1962, A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol. Plant.15:473-497) with 1/2 B5 vitamins, 10 g/L

25 sucrose, 0.05 mg/L NAA, 50 mg/L hygromycin and 2.5 g/L gelrite adjusted to pH 5.8 in magenta boxes. When plantlets were established with well-developed root systems, they were transferred to soil (1 metromix: 1 top soil) and raised in the greenhouse (29/24°C day/night

30 cycle, 50-60% humidity, 12 h photoperiod) until maturity.

#### EXAMPLE 7

Characterization Of Transgenic Rice Plants Expressing

35 Photorhabdus Toxin That Confer Insect Control.

### A. Insect bioassays

Insect bioassays were performed using leaf discs and shown to be highly effective in controlling Southern corn rootworm. *Diabrotica undecimpunctata howardi* eggs are  
 5 obtained from French Ag Research and hatched in petri dishes held at 28.5°C and 40% RH. The aerial parts are sampled from the transgenic plants and placed, singly into inverted petri dishes (100x15mm) containing 15ml of 1.6% aqueous agar in the bottom to provide humidity and  
 10 filter paper in the top to absorb condensation. These preparations are infested with five neonate larvae per dish and held at 28.5°C and 40% RH for 3 days. Mortality and larval weights are recorded. Weight data were transformed using a logarithmic function to correct a  
 15 correlation between the magnitude of the mean and variance.

Table 11

Treatment	Average Survivor Weight in mg <sup>1</sup> (Duncan's Grouping)	Presence TcdA greenhouse-grown plants (number of +/number of plants tested)
GUS Control	0.390 A	-
1553-33	0.170 BCD	++
1553-44	0.167 BCD	+++
1553-62	0.125 CD	+++
1553-41	0.100 D	+++

Note: Means followed by the same letter are not significantly different based on Duncan's multiple range test ( $\alpha=0.05$ ).

20 Insect groups weighing less than 0.1 mg were set to 0.03 mg instead of zero to conduct a more conservative analysis. Weight data were transformed (Log10) for analyses. A single replicate was used on each of three test dates. Plants were  
 25 sampled from magenta boxes.

The results demonstrate that in leaf disc bioassays, several rice events derived by transformation with *tcdA* gene were demonstrated to statistically have a functional affect on corn rootworm neonate.

30

## Claims

1. An isolated nucleic acid of SEQ ID NO: 3 or SEQ ID NO:4.
2. A transgenic monocot cell having a genome comprising  
5 SEQ ID NO:3 or SEQ ID NO:4.
3. A transgenic dicot cell having a genome comprising SEQ ID NO:3 or SEQ ID NO:4.
4. A transgenic plant with a genome comprising a  
nucleic acid of SEQ ID NO: 3 or SEQ ID NO:4 that imparts  
10 insect resistance.
5. A transgenic plant of claim 4 wherein the plant is rice.
6. A transgenic plant of claim 4 wherein the plant is maize.
- 15 7. A transgenic plant of claim 4 wherein the plant is tobacco.

## SEQUENCE LISTING

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<120> Transgenic Plants Expressing Photorhabdus Toxin

<130> 50698

<140>

<141>

<150> US 60/148,356

<151> 1999-08-11

<160> 8

<170> PatentIn Ver. 2.0

<210> 1

<211> 7551

<212> DNA

<213> Photorhabdus luminescens

<220>

<221> CDS

<222> (1)..(7548)

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Gly Phe Asn Cys Leu Thr Asp Ile Ser His Ser Ser Phe Asn Glu Phe	
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Ala Ile Leu Ala Pro Asn Ala Glu Leu Ile Gly Tyr Asn Asn Gln Phe	
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Asn Leu Gln Leu Asp Ile Asn Thr Asp Val Leu Gly Lys Val Phe Leu	
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Val Gln Tyr Cys Gln Ala Leu Ala Gln Leu Glu Met Val Tyr His Ser	
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cac cgt gac acc act tat cca tct aaa gta gaa gct tgg att cct gga 4320  
 His Arg Asp Thr Thr Tyr Pro Ser Lys Val Glu Ala Trp Ile Pro Gly  
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Ser Phe Ser Ile Pro Val Thr Leu Lys Val Ser Thr Asp Asn Ala Leu	
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Gln Glu Pro Gln Leu Gly Lys Gly Phe Tyr Ala Thr Phe Val Ile Pro	
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Asn Arg Trp Leu Lys Tyr Val Trp Ser Pro Ser Gly Tyr Ile Val His	
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Leu Asp Leu Leu Ile Ala Arg Gly Asp His Ala Tyr Arg Gln Leu Glu	
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Ser Leu Arg Ser Ala Asn Thr Leu Thr Asp Leu Phe Leu Pro Gln Ile	
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Ile Tyr Ala Thr Pro Ala Asp Pro Lys Ala Leu Leu Ser Ala Ala Val	
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Asp Ser Tyr Gly Lys Leu Tyr Asp Glu Asn Ile Asn Ala Gly Glu Asn			
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Gln Ala Met Thr Leu Arg Ala Ser Ala Ala Gly Leu Thr Thr Ala Val			
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Gln Ala Ser Arg Leu Ala Gly Ala Ala Ala Asp Leu Val Pro Asn Ile			
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Thr Gly Tyr Val Met Glu Phe Ser Ala Asn Val Met Asn Thr Glu Ala			
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Asp Lys Ile Ser Gln Ser Glu Thr Tyr Arg Arg Arg Arg Gln Glu Trp			
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Glu Ile Gln Arg Asn Asn Ala Glu Ala Glu Leu Lys Gln Ile Asp Ala			
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Thr Ser Leu Lys Thr Gln Gln Glu Gln Thr Gln Ser Gln Leu Ala Phe			
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Leu Gln Arg Lys Phe Ser Asn Gln Ala Leu Tyr Asn Trp Leu Arg Gly			
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Cys Leu Met Ala Glu Gln Ala Tyr Arg Trp Glu Leu Asn Asp Asp Ser			
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Tyr Glu Ile Ala Gln Ala Glu Gln Asp Arg Asn Leu Leu His Glu Lys
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Gly Thr Arg Gln Met Leu Gly Phe Ile Gln Gly Tyr Ser Asp Leu Phe
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Phe Ser Pro Ala Ala Tyr Leu Thr Glu Leu Tyr Arg Glu Ala Lys Asn
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ttg cat gac agc agc tca att tat tac cta gat aaa cgt cgc ccg gat     432
Leu His Asp Ser Ser Ser Ile Tyr Tyr Leu Asp Lys Arg Arg Pro Asp
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acg ctg gct ctc tct aat gaa ttg tgc ctt gcc ggg atc gaa aca aaa     528
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aca gga aaa tca caa gat gaa gtg atg gat atg ttg tca act tat cgt     576
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225 230 235 240	
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His Ile Ser Pro Glu Leu Tyr Asn Leu Leu Ile Glu Glu Ile Pro Glu	
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Ile Thr Thr Ala Gln Leu Met Ser Pro Ser Tyr Leu Ala Arg Tyr Tyr	
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Val Gly Tyr Ser Ser Asp Ile Leu Val Ile Pro Leu Val Asp Gly Val	
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465					470					475					480	
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Lys	Val	Tyr	Arg	Val	Lys	Phe	Tyr	Ile	Asp	Arg	Tyr	Gly	Ile	Ser	Glu	
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Asn	Pro	Asp	Leu	Asn	Leu	Lys	Pro	Asp	Ser	Thr	Gly	Asp	Asp	Gln	Arg	
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Lys	Ala	Val	Leu	Lys	Arg	Ala	Phe	Gln	Val	Asn	Ala	Ser	Glu	Leu	Tyr	
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cag	atg	tta	ttg	atc	act	gat	cgt	aaa	gaa	gac	ggt	ggt	atc	aaa	aat	1776
Gln	Met	Leu	Leu	Ile	Thr	Asp	Arg	Lys	Glu	Asp	Gly	Val	Ile	Lys	Asn	
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Ile	His	Asn	Leu	Thr	Ile	Ala	Glu	Leu	Asn	Ile	Leu	Leu	Val	Ile	Cys	
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Lys	Ile	Val	Glu	Thr	Leu	Leu	Trp	Ile	Thr	Gln	Trp	Leu	Lys	Thr	Gln	
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Ala Tyr Asp Leu Leu Leu	Trp Ile Asp Gln Ile	Gln Pro Ala Gln Ile	
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885	890	895	
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Ile Asp His Asn Tyr Ala	Ala Ala Trp Gln Ala	Ala Ala Ala Ala Leu	
900	905	910	
gct gat cat gct aat cag	gca cag aaa aaa ctg	gat gag acg ttc agt	2784
Ala Asp His Ala Asn Gln	Ala Gln Lys Lys Leu	Asp Glu Thr Phe Ser	
915	920	925	
aag gca tta tgt aac tat	tat att aat gct gtt	gtc gat agt gct gct	2832
Lys Ala Leu Cys Asn Tyr	Tyr Ile Asn Ala Val	Val Val Asp Ser Ala	
930	935	940	

gga gta cgt gat cgt aac ggt tta tat acc tat ttg ctg att gat aat	2880
Gly Val Arg Asp Arg Asn Gly Leu Tyr Thr Tyr Leu Leu Ile Asp Asn	
945 950 955 960	
cag gtt tct gcc gat gtg atc act tca cgt att gca gaa gct atc gcc	2928
Gln Val Ser Ala Asp Val Ile Thr Ser Arg Ile Ala Glu Ala Ile Ala	
965 970 975	
ggt att caa ctg tac gtt aac cgg gct tta aac cga gat gaa ggt cag	2976
Gly Ile Gln Leu Tyr Val Asn Arg Ala Leu Asn Arg Asp Glu Gly Gln	
980 985 990	
ctt gca tcg gac gtt agt acc cgt cag ttc ttc act gac tgg gaa cgt	3024
Leu Ala Ser Asp Val Ser Thr Arg Gln Phe Phe Thr Asp Trp Glu Arg	
995 1000 1005	
tac aat aaa cgt tac agt act tgg gct ggt gtc tct gaa ctg gtc tat	3072
Tyr Asn Lys Arg Tyr Ser Thr Trp Ala Gly Val Ser Glu Leu Val Tyr	
1010 1015 1020	
tat cca gaa aac tat gtt gat ccc act cag cgc att ggg caa acc aaa	3120
Tyr Pro Glu Asn Tyr Val Asp Pro Thr Gln Arg Ile Gly Gln Thr Lys	
1025 1030 1035 1040	
atg atg gat gcg ctg ttg caa tcc atc aac cag agc cag cta aat gcg	3168
Met Met Asp Ala Leu Leu Gln Ser Ile Asn Gln Ser Gln Leu Asn Ala	
1045 1050 1055	
gat acg gtg gaa gat gct ttc aaa act tat ttg acc agc ttt gag cag	3216
Asp Thr Val Glu Asp Ala Phe Lys Thr Tyr Leu Thr Ser Phe Glu Gln	
1060 1065 1070	
gta gca aat ctg aaa gta att agt gct tac cac gat aat gtg aat gtg	3264
Val Ala Asn Leu Lys Val Ile Ser Ala Tyr His Asp Asn Val Asn Val	
1075 1080 1085	
gat caa gga tta act tat ttt atc ggt atc gac caa gca gct ccg ggt	3312
Asp Gln Gly Leu Thr Tyr Phe Ile Gly Ile Asp Gln Ala Ala Pro Gly	
1090 1095 1100	
acg tat tac tgg cgt agt gtt gat cac agc aaa tgt gaa aat ggc aag	3360
Thr Tyr Tyr Trp Arg Ser Val Asp His Ser Lys Cys Glu Asn Gly Lys	
1105 1110 1115 1120	
ttt gcc gct aat gct tgg ggt gag tgg aat aaa att acc tgt gct gtc	3408
Phe Ala Ala Asn Ala Trp Gly Glu Trp Asn Lys Ile Thr Cys Ala Val	
1125 1130 1135	
aat cct tgg aaa aat atc atc cgt ccg gtt gtt tat atg tcc cgc tta	3456
Asn Pro Trp Lys Asn Ile Ile Arg Pro Val Val Tyr Met Ser Arg Leu	
1140 1145 1150	
tat ctg cta tgg ctg gag cag caa tca aag aaa agt gat gat ggt aaa	3504
Tyr Leu Leu Trp Leu Glu Gln Gln Ser Lys Lys Ser Asp Asp Gly Lys	
1155 1160 1165	
acc acg att tat caa tat aac tta aaa ctg gct cat att cgt tac gac	3552
Thr Thr Ile Tyr Gln Tyr Asn Leu Lys Leu Ala His Ile Arg Tyr Asp	
1170 1175 1180	

ggt agt tgg aat aca cca ttt act ttt gat gtg aca gaa aag gta aaa	3600
Gly Ser Trp Asn Thr Pro Phe Thr Phe Asp Val Thr Glu Lys Val Lys	
1185 1190 1195 1200	
aat tac acg tcg agt act gat gct gct gaa tct tta ggg ttg tat tgt	3648
Asn Tyr Thr Ser Ser Thr Asp Ala Ala Glu Ser Leu Gly Leu Tyr Cys	
1205 1210 1215	
act ggt tat caa ggg gaa gac act cta tta gtt atg ttc tat tcg atg	3696
Thr Gly Tyr Gln Gly Glu Asp Thr Leu Leu Val Met Phe Tyr Ser Met	
1220 1225 1230	
cag agt agt tat agc tcc tat acc gat aat aat gcg ccg gtc act ggg	3744
Gln Ser Ser Tyr Ser Ser Tyr Thr Asp Asn Asn Ala Pro Val Thr Gly	
1235 1240 1245	
cta tat att ttc gct gat atg tca tca gac aat atg acg aat gca caa	3792
Leu Tyr Ile Phe Ala Asp Met Ser Ser Asp Asn Met Thr Asn Ala Gln	
1250 1255 1260	
gca act aac tat tgg aat aac agt tat ccg caa ttt gat act gtg atg	3840
Ala Thr Asn Tyr Trp Asn Asn Ser Tyr Pro Gln Phe Asp Thr Val Met	
1265 1270 1275 1280	
gca gat ccg gat agc gac aat aaa aaa gtc ata acc aga aga gtt aat	3888
Ala Asp Pro Asp Ser Asp Asn Lys Lys Val Ile Thr Arg Arg Val Asn	
1285 1290 1295	
aac cgt tat gcg gag gat tat gaa att cct tcc tct gtg aca agt aac	3936
Asn Arg Tyr Ala Glu Asp Tyr Glu Ile Pro Ser Ser Val Thr Ser Asn	
1300 1305 1310	
agt aat tat tct tgg ggt gat cac agt tta acc atg ctt tat ggt ggt	3984
Ser Asn Tyr Ser Trp Gly Asp His Ser Leu Thr Met Leu Tyr Gly Gly	
1315 1320 1325	
agt gtt cct aat att act ttt gaa tcg gcg gca gaa gat tta agg cta	4032
Ser Val Pro Asn Ile Thr Phe Glu Ser Ala Ala Glu Asp Leu Arg Leu	
1330 1335 1340	
tct acc aat atg gca ttg agt att att cat aat gga tat gcg gga acc	4080
Ser Thr Asn Met Ala Leu Ser Ile Ile His Asn Gly Tyr Ala Gly Thr	
1345 1350 1355 1360	
cgc cgt ata caa tgt aat ctt atg aaa caa tac gct tca tta ggt gat	4128
Arg Arg Ile Gln Cys Asn Leu Met Lys Gln Tyr Ala Ser Leu Gly Asp	
1365 1370 1375	
aaa ttt ata att tat gat tca tca ttt gat gat gca aac cgt ttt aat	4176
Lys Phe Ile Ile Tyr Asp Ser Ser Phe Asp Asp Ala Asn Arg Phe Asn	
1380 1385 1390	
ctg gtg cca ttg ttt aaa ttc gga aaa gac gag aac tca gat gat agt	4224
Leu Val Pro Leu Phe Lys Phe Gly Lys Asp Glu Asn Ser Asp Asp Ser	
1395 1400 1405	
att tgt ata tat aat gaa aac cct tcc tct gaa gat aag aag tgg tat	4272
Ile Cys Ile Tyr Asn Glu Asn Pro Ser Ser Glu Asp Lys Lys Trp Tyr	
1410 1415 1420	
ttt tct tcg aaa gat gac aat aaa aca gcg gat tat aat ggt gga act	4320

18

1665	1670	1675	1680	
caa act gtt gtg aaa gtt ttc tta tcc tat ttt ata gag gcg act gga				5088
Gln Thr Val Val Lys Val Phe Leu Ser Tyr Phe Ile Glu Ala Thr Gly				
	1685	1690	1695	
aat aag aac cac tta tgg gta cgt gct aaa tac caa aag gaa acg act				5136
Asn Lys Asn His Leu Trp Val Arg Ala Lys Tyr Gln Lys Glu Thr Thr				
	1700	1705	1710	
gat aag atc ttg ttc gac cgt act gat gag aaa gat ccg cac ggt tgg				5184
Asp Lys Ile Leu Phe Asp Arg Thr Asp Glu Lys Asp Pro His Gly Trp				
	1715	1720	1725	
ttt ctc agc gac gat cac aag acc ttt agt ggt ctc tct tcc gca cag				5232
Phe Leu Ser Asp Asp His Lys Thr Phe Ser Gly Leu Ser Ser Ala Gln				
	1730	1735	1740	
gca tta aag aac gac agt gaa ccg atg gat ttc tct ggc gcc aat gct				5280
Ala Leu Lys Asn Asp Ser Glu Pro Met Asp Phe Ser Gly Ala Asn Ala				
	1745	1750	1755	1760
ctc tat ttc tgg gaa ctg ttc tat tac acg ccg atg atg atg gct cat				5328
Leu Tyr Phe Trp Glu Leu Phe Tyr Tyr Thr Pro Met Met Met Ala His				
	1765	1770	1775	
cgt ttg ttg cag gaa cag aat ttt gat gcg gcg aac cat tgg ttc cgt				5376
Arg Leu Leu Gln Glu Gln Asn Phe Asp Ala Ala Asn His Trp Phe Arg				
	1780	1785	1790	
tat gtc tgg agt cca tcc ggt tat atc gtt gat ggt aaa att gct atc				5424
Tyr Val Trp Ser Pro Ser Gly Tyr Ile Val Asp Gly Lys Ile Ala Ile				
	1795	1800	1805	
tac cac tgg aac gtg cga ccg ctg gaa gaa gac acc agt tgg aat gca				5472
Tyr His Trp Asn Val Arg Pro Leu Glu Glu Asp Thr Ser Trp Asn Ala				
	1810	1815	1820	
caa caa ctg gac tcc acc gat cca gat gct gta gcc caa gat gat ccg				5520
Gln Gln Leu Asp Ser Thr Asp Pro Asp Ala Val Ala Gln Asp Asp Pro				
	1825	1830	1835	1840
atg cac tac aag gtg gct acc ttt atg gcg acg ttg gat ctg cta atg				5568
Met His Tyr Lys Val Ala Thr Phe Met Ala Thr Leu Asp Leu Leu Met				
	1845	1850	1855	
gcc cgt ggt gat gct gct tac cgc cag tta gag cgt gat acg ttg gct				5616
Ala Arg Gly Asp Ala Ala Tyr Arg Gln Leu Glu Arg Asp Thr Leu Ala				
	1860	1865	1870	
gaa gct aaa atg tgg tat aca cag gcg ctt aat ctg ttg ggt gat gag				5664
Glu Ala Lys Met Trp Tyr Thr Gln Ala Leu Asn Leu Leu Gly Asp Glu				
	1875	1880	1885	
cca caa gtg atg ctg agt acg act tgg gct aat cca aca ttg ggt aat				5712
Pro Gln Val Met Leu Ser Thr Thr Trp Ala Asn Pro Thr Leu Gly Asn				
	1890	1895	1900	
gct gct tca aaa acc aca cag cag gtt cgt cag caa gtg ctt acc cag				5760
Ala Ala Ser Lys Thr Thr Gln Gln Val Arg Gln Gln Val Leu Thr Gln				
	1905	1910	1915	1920



ttg cgt ctc aat agc agg gta aaa acc ccg ttg cta gga aca gcc aat	5808
Leu Arg Leu Asn Ser Arg Val Lys Thr Pro Leu Leu Gly Thr Ala Asn	
1925 1930 1935	
tcc ctg acc gct tta ttc ctg ccg cag gaa aat agc aag ctc aaa ggc	5856
Ser Leu Thr Ala Leu Phe Leu Pro Gln Glu Asn Ser Lys Leu Lys Gly	
1940 1945 1950	
tac tgg cgg aca ctg gcg cag cgt atg ttt aat tta cgt cat aat ctg	5904
Tyr Trp Arg Thr Leu Ala Gln Arg Met Phe Asn Leu Arg His Asn Leu	
1955 1960 1965	
tcg att gac ggc cag ccg ctc tcc ttg ccg ctg tat gct aaa ccg gct	5952
Ser Ile Asp Gly Gln Pro Leu Ser Leu Pro Leu Tyr Ala Lys Pro Ala	
1970 1975 1980	
gat cca aaa gct tta ctg agt gcg gcg gtt tca gct tct caa ggg gga	6000
Asp Pro Lys Ala Leu Leu Ser Ala Ala Val Ser Ala Ser Gln Gly Gly	
1985 1990 1995 2000	
gcc gac ttg ccg aag gcg ccg ctg act att cac cgc ttc cct caa atg	6048
Ala Asp Leu Pro Lys Ala Pro Leu Thr Ile His Arg Phe Pro Gln Met	
2005 2010 2015	
cta gaa ggg gca cgg ggc ttg gtt aac cag ctt ata cag ttc ggt agt	6096
Leu Glu Gly Ala Arg Gly Leu Val Asn Gln Leu Ile Gln Phe Gly Ser	
2020 2025 2030	
tca cta ttg ggg tac agt gag cgt cag gat gcg gaa gct atg agt caa	6144
Ser Leu Leu Gly Tyr Ser Glu Arg Gln Asp Ala Glu Ala Met Ser Gln	
2035 2040 2045	
cta ctg caa acc caa gcc agc gag tta ata ctg acc agt att cgt atg	6192
Leu Leu Gln Thr Gln Ala Ser Glu Leu Ile Leu Thr Ser Ile Arg Met	
2050 2055 2060	
cag gat aac caa ttg gca gag ctg gat tcg gaa aaa acc gcc ttg caa	6240
Gln Asp Asn Gln Leu Ala Glu Leu Asp Ser Glu Lys Thr Ala Leu Gln	
2065 2070 2075 2080	
gtc tct tta gct gga gtg caa caa cgg ttt gac agc tat agc caa ctg	6288
Val Ser Leu Ala Gly Val Gln Gln Arg Phe Asp Ser Tyr Ser Gln Leu	
2085 2090 2095	
tat gag gag aac atc aac gca ggt gag cag cga gcg ctg gcg tta cgc	6336
Tyr Glu Glu Asn Ile Asn Ala Gly Glu Gln Arg Ala Leu Ala Leu Arg	
2100 2105 2110	
tca gaa tct gct att gag tct cag gga gcg cag att tcc cgt atg gca	6384
Ser Glu Ser Ala Ile Glu Ser Gln Gly Ala Gln Ile Ser Arg Met Ala	
2115 2120 2125	
ggc gcg ggt gtt gat atg gca cca aat atc ttc ggc ctg gct gat ggc	6432
Gly Ala Gly Val Asp Met Ala Pro Asn Ile Phe Gly Leu Ala Asp Gly	
2130 2135 2140	
ggc atg cat tat ggt gct att gcc tat gcc atc gct gac ggt att gag	6480
Gly Met His Tyr Gly Ala Ile Ala Tyr Ala Ile Ala Asp Gly Ile Glu	
2145 2150 2155 2160	

ttg agt gct tct gcc aag atg gtt gat gcg gag aaa gtt gct cag tcg Leu Ser Ala Ser Ala Lys Met Val Asp Ala Glu Lys Val Ala Gln Ser 2165 2170 2175	6528
gaa ata tat cgc cgt cgc cgt caa gaa tgg aaa att cag cgt gac aac Glu Ile Tyr Arg Arg Arg Gln Glu Trp Lys Ile Gln Arg Asp Asn 2180 2185 2190	6576
gca caa gcg gag att aac cag tta aac gcg caa ctg gaa tca ctg tct Ala Gln Ala Glu Ile Asn Gln Leu Asn Ala Gln Leu Glu Ser Leu Ser 2195 2200 2205	6624
att cgc cgt gaa gcc gct gaa atg caa aaa gag tac ctg aaa acc cag Ile Arg Arg Glu Ala Ala Glu Met Gln Lys Glu Tyr Leu Lys Thr Gln 2210 2215 2220	6672
caa gct cag gcg cag gca caa ctt act ttc tta aga agc aaa ttc agt Gln Ala Gln Ala Gln Ala Gln Leu Thr Phe Leu Arg Ser Lys Phe Ser 2225 2230 2235 2240	6720
aat caa gcg tta tat agt tgg tta cga ggg cgt ttg tca ggt att tat Asn Gln Ala Leu Tyr Ser Trp Leu Arg Gly Arg Leu Ser Gly Ile Tyr 2245 2250 2255	6768
ttc cag ttc tat gac ttg gcc gta tca cgt tgc ctg atg gca gag caa Phe Gln Phe Tyr Asp Leu Ala Val Ser Arg Cys Leu Met Ala Glu Gln 2260 2265 2270	6816
tcc tat caa tgg gaa gct aat gat aat tcc att agc ttt gtc aaa ccg Ser Tyr Gln Trp Glu Ala Asn Asp Asn Ser Ile Ser Phe Val Lys Pro 2275 2280 2285	6864
ggt gca tgg caa gga act tac gcc ggc tta ttg tgt gga gaa gct ttg Gly Ala Trp Gln Gly Thr Tyr Ala Gly Leu Leu Cys Gly Glu Ala Leu 2290 2295 2300	6912
ata caa aat ctg gca caa atg gaa gag gca tat ctg aaa tgg gaa tct Ile Gln Asn Leu Ala Gln Met Glu Glu Ala Tyr Leu Lys Trp Glu Ser 2305 2310 2315 2320	6960
cgc gct ttg gaa gta gaa cgc acg gtt tca ttg gca gtg gtt tat gat Arg Ala Leu Glu Val Glu Arg Thr Val Ser Leu Ala Val Val Tyr Asp 2325 2330 2335	7008
tca ctg gaa ggt aat gat cgt ttt aat tta gcg gaa caa ata cct gca Ser Leu Glu Gly Asn Asp Arg Phe Asn Leu Ala Glu Gln Ile Pro Ala 2340 2345 2350	7056
tta ttg gat aag ggg gag gga aca gca gga act aaa gaa aat ggg tta Leu Leu Asp Lys Gly Glu Gly Thr Ala Gly Thr Lys Glu Asn Gly Leu 2355 2360 2365	7104
tca ttg gct aat gct atc ctg tca gct tcg gtc aaa ttg tcc gac ttg Ser Leu Ala Asn Ala Ile Leu Ser Ala Ser Val Lys Leu Ser Asp Leu 2370 2375 2380	7152
aaa ctg gga acg gat tat cca gac agt atc gtt ggt agc aac aag gtt Lys Leu Gly Thr Asp Tyr Pro Asp Ser Ile Val Gly Ser Asn Lys Val 2385 2390 2395 2400	7200
cgt cgt att aag caa atc agt gtt tcg cta cct gca ttg gtt ggg cct	7248

Arg Arg Ile Lys Gln Ile Ser Val Ser Leu Pro Ala Leu Val Gly Pro  
 2405 2410 2415

tat cag gat gtt cag gct atg ctc agc tat ggt ggc agt act caa ttg 7296  
 Tyr Gln Asp Val Gln Ala Met Leu Ser Tyr Gly Gly Ser Thr Gln Leu  
 2420 2425 2430

ccg aaa ggt tgt tca gcg ttg gct gtg tct cat ggt acc aat gat agt 7344  
 Pro Lys Gly Cys Ser Ala Leu Ala Val Ser His Gly Thr Asn Asp Ser  
 2435 2440 2445

ggt cag ttc cag ttg gat ttc aat gac ggc aaa tac ctg cca ttt gaa 7392  
 Gly Gln Phe Gln Leu Asp Phe Asn Asp Gly Lys Tyr Leu Pro Phe Glu  
 2450 2455 2460

ggt att gct ctt gat gat cag ggt aca ctg aat ctt caa ttt ccg aat 7440  
 Gly Ile Ala Leu Asp Asp Gln Gly Thr Leu Asn Leu Gln Phe Pro Asn  
 2465 2470 2475 2480

gct acc gac aag cag aaa gca ata ttg caa act atg agc gat att att 7488  
 Ala Thr Asp Lys Gln Lys Ala Ile Leu Gln Thr Met Ser Asp Ile Ile  
 2485 2490 2495

ttg cat att cgt tat acc atc cgt taa 7515  
 Leu His Ile Arg Tyr Thr Ile Arg  
 2500

&lt;210&gt; 3

&lt;211&gt; 7577

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (3)..(7553)

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence:hemicot tcdA

&lt;400&gt; 3

cc atg gct aac gag tcc gtc aag gag atc cca gac gtc ctc aag tcc 47  
 Met Ala Asn Glu Ser Val Lys Glu Ile Pro Asp Val Leu Lys Ser  
 1 5 10 15

caa tgc ggt ttc aac tgc ctc act gac atc tcc cac agc tcc ttc aac 95  
 Gln Cys Gly Phe Asn Cys Leu Thr Asp Ile Ser His Ser Ser Phe Asn  
 20 25 30

gag ttc aga caa caa gtc tct gag cac ctc tcc tgg tcc gag acc cat 143  
 Glu Phe Arg Gln Gln Val Ser Glu His Leu Ser Trp Ser Glu Thr His  
 35 40 45

gac ctc tac cat gac gct cag caa gct cag aag gac aac agg ctc tac 191  
 Asp Leu Tyr His Asp Ala Gln Gln Ala Gln Lys Asp Asn Arg Leu Tyr  
 50 55 60

gag gct agg atc ctc aag agg gct aac cca caa ctc cag aac gct gtc 239  
 Glu Ala Arg Ile Leu Lys Arg Ala Asn Pro Gln Leu Gln Asn Ala Val  
 65 70 75

cac ctc gcc atc ttg gct cca aac gct gag ttg att ggt tac aac aac	287
His Leu Ala Ile Leu Ala Pro Asn Ala Glu Leu Ile Gly Tyr Asn Asn	
80 85 90 95	
cag ttc tct ggc aga gct agc cag tac gtg gct cct ggt aca gtc tcc	335
Gln Phe Ser Gly Arg Ala Ser Gln Tyr Val Ala Pro Gly Thr Val Ser	
100 105 110	
tcc atg ttc agc cca gcc gct tac ctc act gag ttg tac cgc gag gct	383
Ser Met Phe Ser Pro Ala Ala Tyr Leu Thr Glu Leu Tyr Arg Glu Ala	
115 120 125	
agg aac ctt cat gct tct gac tcc gtc tac tac ttg gac aca cgc aga	431
Arg Asn Leu His Ala Ser Asp Ser Val Tyr Tyr Leu Asp Thr Arg Arg	
130 135 140	
cca gac ctc aag agc atg gcc ctc agc caa cag aac atg gac att gag	479
Pro Asp Leu Lys Ser Met Ala Leu Ser Gln Gln Asn Met Asp Ile Glu	
145 150 155	
ttg tcc acc ctc tcc ttg agc aac gag ctt ctc ttg gag tcc atc aag	527
Leu Ser Thr Leu Ser Leu Ser Asn Glu Leu Leu Glu Ser Ile Lys	
160 165 170 175	
act gag agc aag ttg gag aac tac acc aag gtc atg gag atg ctc tcc	575
Thr Glu Ser Lys Leu Glu Asn Tyr Thr Lys Val Met Glu Met Leu Ser	
180 185 190	
acc ttc aga cca agc ggt gca act cca tac cat gat gcc tac gag aac	623
Thr Phe Arg Pro Ser Gly Ala Thr Pro Tyr His Asp Ala Tyr Glu Asn	
195 200 205	
gtc agg gag gtc atc caa ctt caa gac cct ggt ctt gag caa ctc aac	671
Val Arg Glu Val Ile Gln Leu Gln Asp Pro Gly Leu Glu Gln Leu Asn	
210 215 220	
gct tct cca gcc att gct ggt ttg atg cac cag gca tcc ttg ctc ggt	719
Ala Ser Pro Ala Ile Ala Gly Leu Met His Gln Ala Ser Leu Leu Gly	
225 230 235	
atc aac gcc tcc atc tct cct gag ttg ttc aac atc ttg act gag gag	767
Ile Asn Ala Ser Ile Ser Pro Glu Leu Phe Asn Ile Leu Thr Glu Glu	
240 245 250 255	
atc act gag ggc aac gct gag gag ttg tac aag aag aac ttc ggc aac	815
Ile Thr Glu Gly Asn Ala Glu Glu Leu Tyr Lys Lys Asn Phe Gly Asn	
260 265 270	
att gag cca gcc tct ctt gca atg cct gag tac ctc aag agg tac tac	863
Ile Glu Pro Ala Ser Leu Ala Met Pro Glu Tyr Leu Lys Arg Tyr Tyr	
275 280 285	
aac ttg tct gat gag gag ctt tct caa ttc att ggc aag gct tcc aac	911
Asn Leu Ser Asp Glu Glu Leu Ser Gln Phe Ile Gly Lys Ala Ser Asn	
290 295 300	
ttc ggt caa cag gag tac agc aac aac cag ctc atc act cca gtt gtg	959
Phe Gly Gln Gln Glu Tyr Ser Asn Asn Gln Leu Ile Thr Pro Val Val	
305 310 315	

aac tcc tct gat ggc act gtg aag gtc tac cgc atc aca cgt gag tac	1007
Asn Ser Ser Asp Gly Thr Val Lys Val Tyr Arg Ile Thr Arg Glu Tyr	
320 325 330 335	
acc aca aac gcc tac caa atg gat gtt gag ttg ttc cca ttc ggt ggt	1055
Thr Thr Asn Ala Tyr Gln Met Asp Val Glu Leu Phe Pro Phe Gly Gly	
340 345 350	
gag aac tac aga ctt gac tac aag ttc aag aac ttc tac aac gcc tcc	1103
Glu Asn Tyr Arg Leu Asp Tyr Lys Phe Lys Asn Phe Tyr Asn Ala Ser	
355 360 365	
tac ctc tcc atc aag ttg aac gac aag agg gag ctt gtc agg act gag	1151
Tyr Leu Ser Ile Lys Leu Asn Asp Lys Arg Glu Leu Val Arg Thr Glu	
370 375 380	
ggt gct cct caa gtg aac att gag tac tct gcc aac atc acc ctc aac	1199
Gly Ala Pro Gln Val Asn Ile Glu Tyr Ser Ala Asn Ile Thr Leu Asn	
385 390 395	
aca gct gac atc tct caa cca ttc gag att ggt ttg acc aga gtc ctt	1247
Thr Ala Asp Ile Ser Gln Pro Phe Glu Ile Gly Leu Thr Arg Val Leu	
400 405 410 415	
ccc tct ggc tcc tgg gcc tac gct gca gcc aag ttc act gtt gag gag	1295
Pro Ser Gly Ser Trp Ala Tyr Ala Ala Lys Phe Thr Val Glu Glu	
420 425 430	
tac aac cag tac tct ttc ctc ttg aag ctc aac aag gca att cgt ctc	1343
Tyr Asn Gln Tyr Ser Phe Leu Leu Lys Leu Asn Lys Ala Ile Arg Leu	
435 440 445	
agc aga gcc act gag ttg tct ccc acc atc ttg gag ggc att gtg agg	1391
Ser Arg Ala Thr Glu Leu Ser Pro Thr Ile Leu Glu Gly Ile Val Arg	
450 455 460	
tct gtc aac ctt caa ctt gac atc aac act gat gtg ctt ggc aag gtc	1439
Ser Val Asn Leu Gln Leu Asp Ile Asn Thr Asp Val Leu Gly Lys Val	
465 470 475	
ttc ctc acc aag tac tac atg caa cgc tac gcc atc cat gct gag act	1487
Phe Leu Thr Lys Tyr Tyr Met Gln Arg Tyr Ala Ile His Ala Glu Thr	
480 485 490 495	
gca ctc atc ctc tgc aac gca ccc atc tct caa cgc tcc tac gac aac	1535
Ala Leu Ile Leu Cys Asn Ala Pro Ile Ser Gln Arg Ser Tyr Asp Asn	
500 505 510	
cag cct tcc cag ttc gac agg ctc ttc aac act cct ctc ttg aac ggc	1583
Gln Pro Ser Gln Phe Asp Arg Leu Phe Asn Thr Pro Leu Leu Asn Gly	
515 520 525	
cag tac ttc tcc act ggt gat gag gag att gac ctc aac tct ggc tcc	1631
Gln Tyr Phe Ser Thr Gly Asp Glu Glu Ile Asp Leu Asn Ser Gly Ser	
530 535 540	
aca ggt gac tgg aga aag acc atc ttg aag agg gcc ttc aac att gat	1679
Thr Gly Asp Trp Arg Lys Thr Ile Leu Lys Arg Ala Phe Asn Ile Asp	
545 550 555	
gat gtc tct ctc ttc cgt ctc ttg aag atc aca gat cac gac aac aag	1727

Asp Val Ser Leu Phe Arg Leu Leu Lys Ile Thr Asp His Asp Asn Lys	
560 565 570 575	
gat ggc aag atc aag aac aac ttg aag aac ctt tcc aac ctc tac att	1775
Asp Gly Lys Ile Lys Asn Asn Leu Lys Asn Leu Ser Asn Leu Tyr Ile	
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Gly Lys Leu Leu Ala Asp Ile His Gln Leu Thr Ile Asp Glu Leu Asp	
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Leu Leu Leu Ile Ala Val Gly Glu Gly Lys Thr Asn Leu Ser Ala Ile	
610 615 620	
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Ser Asp Lys Gln Leu Ala Thr Leu Ile Arg Lys Leu Asn Thr Ile Thr	
625 630 635	
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Ser Trp Leu His Thr Gln Lys Trp Ser Val Phe Gln Leu Phe Ile Met	
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Leu Asp Thr Val Tyr His Gly Leu Gln Gly Phe Asp Lys Asp Lys Ala	
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Ser Ser Glu Asn Val Ala His Ser Val Leu Leu Trp Ala Asp Lys Leu	
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Gln Pro Gly Asp Gly Ala Met Thr Ala Glu Lys Phe Trp Asp Trp Leu	
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Gln Trp Val Asn Val Ala Gln Gln Leu Asn Val Ala Pro Gln Gly Val	880	885	890	895
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Ser Ala Leu Val Gly Leu Asp Tyr Ile Gln Ser Met Lys Glu Thr Pro	900	905	910	
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Thr Tyr Ala Gln Trp Glu Asn Ala Ala Gly Val Leu Thr Ala Gly Leu	915	920	925	
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Ala Ala Ile Lys Ser Arg Asp Asp Leu Tyr Gln Tyr Leu Leu Ile Asp	960	965	970	975
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           20             25             30

acc ttc cgt gag aag acc aga ggc atg gtc aac tgg ggt gag gcc aag      143
Thr Phe Arg Glu Lys Thr Arg Gly Met Val Asn Trp Gly Glu Ala Lys
           35             40             45

agg atc tac gag att gct caa gct gag caa gac agg aac ctc ctt cat      191
Arg Ile Tyr Glu Ile Ala Gln Ala Glu Gln Asp Arg Asn Leu Leu His
           50             55             60

gag aag agg atc ttc gcc tac gct aac cca ttg ctc aag aac gct gtc      239
Glu Lys Arg Ile Phe Ala Tyr Ala Asn Pro Leu Leu Lys Asn Ala Val
           65             70             75

agg ctt ggt acc agg caa atg ttg ggt ttc atc caa ggt tac tct gac      287
Arg Leu Gly Thr Arg Gln Met Leu Gly Phe Ile Gln Gly Tyr Ser Asp
           80             85             90             95

ttg ttc ggc aac agg gct gac aac tac gca gct cct ggt tct gtt gct      335
Leu Phe Gly Asn Arg Ala Asp Asn Tyr Ala Ala Pro Gly Ser Val Ala
           100            105            110

agc atg ttc agc cca gct gcc tac ctc act gag ttg tac cgt gag gcc      383
Ser Met Phe Ser Pro Ala Ala Tyr Leu Thr Glu Leu Tyr Arg Glu Ala
           115            120            125

aag aac ctc cat gac agc tcc agc atc tac tac ctt gac aag agg cgc      431
Lys Asn Leu His Asp Ser Ser Ser Ile Tyr Tyr Leu Asp Lys Arg Arg
           130            135            140

cca gac ctt gct tcc ttg atg ctc tcc cag aag aac atg gat gag gag      479
Pro Asp Leu Ala Ser Leu Met Leu Ser Gln Lys Asn Met Asp Glu Glu
           145            150            155

atc agc acc ttg gct ctc tcc aac gag ctt tgc ttg gct ggc att gag      527
Ile Ser Thr Leu Ala Leu Ser Asn Glu Leu Cys Leu Ala Gly Ile Glu
           160            165            170            175

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acc aag act ggc aag tcc caa gat gag gtc atg gac atg ctc tcc acc	575
Thr Lys Thr Gly Lys Ser Gln Asp Glu Val Met Asp Met Leu Ser Thr	
180 185 190	
tac cgc ctc tct ggt gag act cca tac cac cat gct tac gag act gtc	623
Tyr Arg Leu Ser Gly Glu Thr Pro Tyr His His Ala Tyr Glu Thr Val	
195 200 205	
agg gag att gtc cat gag agg gac cca ggt ttc cgc cac ctc tcc caa	671
Arg Glu Ile Val His Glu Arg Asp Pro Gly Phe Arg His Leu Ser Gln	
210 215 220	
gct ccc att gtg gct gcc aag ttg gac cca gtc acc ctc ttg ggc atc	719
Ala Pro Ile Val Ala Ala Lys Leu Asp Pro Val Thr Leu Leu Gly Ile	
225 230 235	
tcc agc cac atc agc cca gag ttg tac aac ctt ctc att gag gag atc	767
Ser Ser His Ile Ser Pro Glu Leu Tyr Asn Leu Leu Ile Glu Glu Ile	
240 245 250 255	
cca gag aag gat gag gca gct ttg gac acc ctc tac aag acc aac ttc	815
Pro Glu Lys Asp Glu Ala Ala Leu Asp Thr Leu Tyr Lys Thr Asn Phe	
260 265 270	
ggt gac atc acc act gct caa ctc atg agc cca tcc tac ttg gcc agg	863
Gly Asp Ile Thr Thr Ala Gln Leu Met Ser Pro Ser Tyr Leu Ala Arg	
275 280 285	
tac tac ggt gtc tct cca gag gac att gct tac gtc acc aca agc ctc	911
Tyr Tyr Gly Val Ser Pro Glu Asp Ile Ala Tyr Val Thr Thr Ser Leu	
290 295 300	
tcc cat gtg ggt tac tcc tct gac atc ctt gtc atc cca ctc gtg gat	959
Ser His Val Gly Tyr Ser Ser Asp Ile Leu Val Ile Pro Leu Val Asp	
305 310 315	
ggt gtg ggc aag atg gag gtt gtc agg gtc acc agg act cca tct gac	1007
Gly Val Gly Lys Met Glu Val Val Arg Val Thr Arg Thr Pro Ser Asp	
320 325 330 335	
aac tac acc tcc cag acc aac tac att gag ttg tac cca caa ggt ggt	1055
Asn Tyr Thr Ser Gln Thr Asn Tyr Ile Glu Leu Tyr Pro Gln Gly Gly	
340 345 350	
gac aac tac ctc atc aag tac aac ctc tcc aac tct ttc ggt ttg gat	1103
Asp Asn Tyr Leu Ile Lys Tyr Asn Leu Ser Asn Ser Phe Gly Leu Asp	
355 360 365	
gac ttc tac ctc cag tac aag gat ggt tct gct gac tgg act gag att	1151
Asp Phe Tyr Leu Gln Tyr Lys Asp Gly Ser Ala Asp Trp Thr Glu Ile	
370 375 380	
gct cac aac cca tac cca gac atg gtc atc aac cag aag tac gag tcc	1199
Ala His Asn Pro Tyr Pro Asp Met Val Ile Asn Gln Lys Tyr Glu Ser	
385 390 395	
caa gcc acc atc aag aga tct gac tct gac aac atc ctc tcc att ggt	1247
Gln Ala Thr Ile Lys Arg Ser Asp Ser Asp Asn Ile Leu Ser Ile Gly	
400 405 410 415	
ctc caa agg tgg cac tct ggt tcc tac aac ttc gct gct gcc aac ttc	1295

Leu Gln Arg Trp His Ser Gly Ser Tyr Asn Phe Ala Ala Ala Asn Phe	
420 425 430	
aag att gac caa tac tct cca aag gct ttc ctc ttg aag atg aac aag	1343
Lys Ile Asp Gln Tyr Ser Pro Lys Ala Phe Leu Leu Lys Met Asn Lys	
435 440 445	
gcc atc agg ctc ttg aag gcc act ggt ctc tcc ttc gcc acc ctt gag	1391
Ala Ile Arg Leu Leu Lys Ala Thr Gly Leu Ser Phe Ala Thr Leu Glu	
450 455 460	
agg att gtg gac tct gtc aac tcc acc aag tcc atc act gtg gag gtc	1439
Arg Ile Val Asp Ser Val Asn Ser Thr Lys Ser Ile Thr Val Glu Val	
465 470 475	
ctc aac aag gtc tac aga gtc aag ttc tac att gac cgc tac ggc atc	1487
Leu Asn Lys Val Tyr Arg Val Lys Phe Tyr Ile Asp Arg Tyr Gly Ile	
480 485 490 495	
tct gag gag act gct gcc atc ctt gcc aac atc aac atc tcc cag caa	1535
Ser Glu Glu Thr Ala Ala Ile Leu Ala Asn Ile Asn Ile Ser Gln Gln	
500 505 510	
gct gtc ggc aac cag ctc tcc caa ttc gag caa ctc ttc aac cac cct	1583
Ala Val Gly Asn Gln Leu Ser Gln Phe Glu Gln Leu Phe Asn His Pro	
515 520 525	
cca ctc aac ggc atc cgc tac gag atc agc gag gac aac tcc aag cac	1631
Pro Leu Asn Gly Ile Arg Tyr Glu Ile Ser Glu Asp Asn Ser Lys His	
530 535 540	
ctc cca aac cca gac ctc aac ctc aag cca gac tcc act ggt gat gac	1679
Leu Pro Asn Pro Asp Leu Asn Leu Lys Pro Asp Ser Thr Gly Asp Asp	
545 550 555	
caa agg aag gct gtc ctc aag agg gct ttc caa gtc aac gct tct gag	1727
Gln Arg Lys Ala Val Leu Lys Arg Ala Phe Gln Val Asn Ala Ser Glu	
560 565 570 575	
ctt tac caa atg ctc ttg atc act gac agg aag gag gat ggt gtc atc	1775
Leu Tyr Gln Met Leu Leu Ile Thr Asp Arg Lys Glu Asp Gly Val Ile	
580 585 590	
aag aac aac ttg gag aac ctc tct gac ctc tac ctt gtc tcc ctc ttg	1823
Lys Asn Asn Leu Glu Asn Leu Ser Asp Leu Tyr Leu Val Ser Leu Leu	
595 600 605	
gcc caa atc cac aac ttg acc att gct gag ttg aac atc ctc ttg gtc	1871
Ala Gln Ile His Asn Leu Thr Ile Ala Glu Leu Asn Ile Leu Leu Val	
610 615 620	
atc tgc ggt tac ggt gac acc aac atc tac caa atc act gac gac aac	1919
Ile Cys Gly Tyr Gly Asp Thr Asn Ile Tyr Gln Ile Thr Asp Asp Asn	
625 630 635	
ctt gcc aag att gtg gag acc ctc ttg tgg atc acc caa tgg ctc aag	1967
Leu Ala Lys Ile Val Glu Thr Leu Leu Trp Ile Thr Gln Trp Leu Lys	
640 645 650 655	
acc cag aag tgg act gtc aca gac ctc ttc ctc atg acc act gcc acc	2015
Thr Gln Lys Trp Thr Val Thr Asp Leu Phe Leu Met Thr Thr Ala Thr	



660										665										670									
tac	tcc	acc	act	ctc	act	cca	gag	att	tcc	aac	ctc	act	gcc	acc	ctc														
Tyr	Ser	Thr	Thr	Leu	Thr	Pro	Glu	Ile	Ser	Asn	Leu	Thr	Ala	Thr	Leu														
			675					680					685																2063
agc	tcc	acc	ctc	cac	ggc	aag	gag	tcc	ctc	att	ggt	gag	gac	ctc	aag														
Ser	Ser	Thr	Leu	His	Gly	Lys	Glu	Ser	Leu	Ile	Gly	Glu	Asp	Leu	Lys														
			690					695					700																2111
agg	gca	atg	gct	cca	tgc	ttc	acc	tct	gct	ctc	cac	ctc	acc	tcc	caa														
Arg	Ala	Met	Ala	Pro	Cys	Phe	Thr	Ser	Ala	Leu	His	Leu	Thr	Ser	Gln														
			705					710					715																2159
gag	gtg	gct	tac	gac	ctc	ctt	ctc	tgg	att	gac	caa	atc	caa	cca	gct														
Glu	Val	Ala	Tyr	Asp	Leu	Leu	Leu	Trp	Ile	Asp	Gln	Ile	Gln	Pro	Ala														
																													2207
caa	atc	act	gtg	gat	ggc	ttc	tgg	gag	gag	gtc	caa	acc	act	cca	acc														
Gln	Ile	Thr	Val	Asp	Gly	Phe	Trp	Glu	Glu	Val	Gln	Thr	Thr	Pro	Thr														
																													2255
tcc	ctc	aag	gtc	atc	acc	ttc	gct	caa	gtc	ttg	gct	caa	ctc	tcc	ctc														
Ser	Leu	Lys	Val	Ile	Thr	Phe	Ala	Gln	Val	Leu	Ala	Gln	Leu	Ser	Leu														
																													2303
atc	tac	aga	agg	att	ggc	ctc	tct	gag	act	gag	ttg	tcc	ctc	att	gtc														
Ile	Tyr	Arg	Arg	Ile	Gly	Leu	Ser	Glu	Thr	Glu	Leu	Ser	Leu	Ile	Val														
																													2351
acc	caa	tcc	agc	ctc	ttg	gtc	gct	ggc	aag	tcc	atc	ctt	gat	cat	ggc														
Thr	Gln	Ser	Ser	Leu	Leu	Val	Ala	Gly	Lys	Ser	Ile	Leu	Asp	His	Gly														
																													2399
ctc	ttg	acc	ctc	atg	gct	ctt	gag	ggc	ttc	cac	acc	tgg	gtc	aac	ggc														
Leu	Leu	Thr	Leu	Met	Ala	Leu	Glu	Gly	Phe	His	Thr	Trp	Val	Asn	Gly														
																													2447
ttg	ggc	caa	cat	gct	tcc	ctc	atc	ttg	gct	gca	ctc	aag	gat	ggc	gct														
Leu	Gly	Gln	His	Ala	Ser	Leu	Ile	Leu	Ala	Ala	Leu	Lys	Asp	Gly	Ala														
																													2495
ctc	acc	gtc	acc	gat	gtg	gct	caa	gcc	atg	aac	aag	gag	gag	tcc	ctc														
Leu	Thr	Val	Thr	Asp	Val	Ala	Gln	Ala	Met	Asn	Lys	Glu	Glu	Ser	Leu														
																													2543
ttg	caa	atg	gct	gcc	aac	cag	gtg	gag	aag	gac	ctc	acc	aag	ctc	acc														
Leu	Gln	Met	Ala	Ala	Asn	Gln	Val	Glu	Lys	Asp	Leu	Thr	Lys	Leu	Thr														
																													2591
tcc	tgg	acc	caa	atc	gat	gcc	atc	ctc	caa	tgg	ctc	caa	atg	tcc	tct														
Ser	Trp	Thr	Gln	Ile	Asp	Ala	Ile	Leu	Gln	Trp	Leu	Gln	Met	Ser	Ser														
																													2639
gct	ctt	gct	gtc	agc	cca	ttg	gac	ctt	gct	ggc	atg	atg	gct	ctc	aag														
Ala	Leu	Ala	Val	Ser	Pro	Leu	Asp	Leu	Ala	Gly	Met	Met	Ala	Leu	Lys														
																													2687
tac	ggc	att	gat	cac	aac	tac	gct	gcc	tgg	caa	gca	gct	gcc	gct	gcc														
Tyr	Gly	Ile	Asp	His	Asn	Tyr	Ala	Ala	Trp	Gln	Ala	Ala	Ala	Ala	Ala														
																													2735

ctc atg gct gac cat gcc aac cag gct cag aag aag ttg gat gag acc	2783
Leu Met Ala Asp His Ala Asn Gln Ala Gln Lys Lys Leu Asp Glu Thr	
915 920 925	
ttc tcc aag gct ctc tgc aac tac tac atc aac gcc gtg gtt gac tct	2831
Phe Ser Lys Ala Leu Cys Asn Tyr Tyr Ile Asn Ala Val Val Asp Ser	
930 935 940	
gct gcc ggt gtc agg gac agg aac ggt ctc tac acc tac ctc ttg att	2879
Ala Ala Gly Val Arg Asp Arg Asn Gly Leu Tyr Thr Tyr Leu Leu Ile	
945 950 955	
gac aac cag gtc tct gct gat gtc atc acc tcc aga att gct gag gcc	2927
Asp Asn Gln Val Ser Ala Asp Val Ile Thr Ser Arg Ile Ala Glu Ala	
960 965 970 975	
att gct ggc atc caa ctc tac gtc aac agg gct ctc aac agg gat gag	2975
Ile Ala Gly Ile Gln Leu Tyr Val Asn Arg Ala Leu Asn Arg Asp Glu	
980 985 990	
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Gly Gln Leu Ala Ser Asp Val Ser Thr Arg Gln Phe Phe Thr Asp Trp	
995 1000 1005	
gag agg tac aac aag agg tac tcc acc tgg gct ggt gtc tct gag ttg	3071
Glu Arg Tyr Asn Lys Arg Tyr Ser Thr Trp Ala Gly Val Ser Glu Leu	
1010 1015 1020	
gtc tac tac cca gag aac tac gtg gac cca acc caa agg att ggt cag	3119
Val Tyr Tyr Pro Glu Asn Tyr Val Asp Pro Thr Gln Arg Ile Gly Gln	
1025 1030 1035	
acc aag atg atg gat gct ttg ctc caa tcc atc aac cag tcc caa ctc	3167
Thr Lys Met Met Asp Ala Leu Leu Gln Ser Ile Asn Gln Ser Gln Leu	
1040 1045 1050 1055	
aac gct gac act gtg gag gat gct ttc aag acc tac ctc acc tcc ttc	3215
Asn Ala Asp Thr Val Glu Asp Ala Phe Lys Thr Tyr Leu Thr Ser Phe	
1060 1065 1070	
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Glu Gln Val Ala Asn Leu Lys Val Ile Ser Ala Tyr His Asp Asn Val	
1075 1080 1085	
aac gtg gac caa ggt ctc acc tac ttc att ggc att gac caa gcc gct	3311
Asn Val Asp Gln Gly Leu Thr Tyr Phe Ile Gly Ile Asp Gln Ala Ala	
1090 1095 1100	
cct ggc acc tac tac tgg agg tct gtg gac cac tcc aag tgc gag aac	3359
Pro Gly Thr Tyr Tyr Trp Arg Ser Val Asp His Ser Lys Cys Glu Asn	
1105 1110 1115	
ggc aag ttc gct gcc aac gct tgg ggt gag tgg aac aag atc acc tgc	3407
Gly Lys Phe Ala Ala Asn Ala Trp Gly Glu Trp Asn Lys Ile Thr Cys	
1120 1125 1130 1135	
gct gtc aac cct tgg aag aac atc atc agg cca gtg gtc tac atg tcc	3455
Ala Val Asn Pro Trp Lys Asn Ile Ile Arg Pro Val Val Tyr Met Ser	
1140 1145 1150	

aga ctc tac ttg ctc tgg ctt gag caa cag tcc aag aag tct gat gac	3503
Arg Leu Tyr Leu Leu Trp Leu Glu Gln Gln Ser Lys Lys Ser Asp Asp	
1155 1160 1165	
ggc aag aca act atc tac cag tac aac ctc aag ttg gct cac atc cgc	3551
Gly Lys Thr Thr Ile Tyr Gln Tyr Asn Leu Lys Leu Ala His Ile Arg	
1170 1175 1180	
tac gat ggt tcc tgg aac act cca ttc acc ttc gat gtc act gag aag	3599
Tyr Asp Gly Ser Trp Asn Thr Pro Phe Thr Phe Asp Val Thr Glu Lys	
1185 1190 1195	
gtc aag aac tac acc tcc agc act gat gca gct gag tcc ctt ggt ctc	3647
Val Lys Asn Tyr Thr Ser Ser Thr Asp Ala Ala Glu Ser Leu Gly Leu	
1200 1205 1210 1215	
tac tgc act ggt tac caa ggt gag gac acc ctc ttg gtc atg ttc tac	3695
Tyr Cys Thr Gly Tyr Gln Gly Glu Asp Thr Leu Leu Val Met Phe Tyr	
1220 1225 1230	
tcc atg caa tcc agc tac tcc agc tac act gac aac aac gct cca gtc	3743
Ser Met Gln Ser Ser Tyr Ser Ser Tyr Thr Asp Asn Asn Ala Pro Val	
1235 1240 1245	
act ggt ctc tac atc ttc gct gac atg tcc tct gac aac atg acc aac	3791
Thr Gly Leu Tyr Ile Phe Ala Asp Met Ser Ser Asp Asn Met Thr Asn	
1250 1255 1260	
gct caa gcc acc aac tac tgg aac aac tcc tac cca caa ttc gac act	3839
Ala Gln Ala Thr Asn Tyr Trp Asn Asn Ser Tyr Pro Gln Phe Asp Thr	
1265 1270 1275	
gtc atg gct gac cca gac tct gac aac aag aag gtc atc acc agg cgt	3887
Val Met Ala Asp Pro Asp Ser Asp Asn Lys Lys Val Ile Thr Arg Arg	
1280 1285 1290 1295	
gtc aac aac cgc tac gct gag gac tac gag atc cca agc tct gtc acc	3935
Val Asn Asn Arg Tyr Ala Glu Asp Tyr Glu Ile Pro Ser Ser Val Thr	
1300 1305 1310	
tcc aac agc aac tac tcc tgg ggt gac cac tcc ctc acc atg ctc tac	3983
Ser Asn Ser Asn Tyr Ser Trp Gly Asp His Ser Leu Thr Met Leu Tyr	
1315 1320 1325	
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Gly Gly Ser Val Pro Asn Ile Thr Phe Glu Ser Ala Ala Glu Asp Leu	
1330 1335 1340	
agg ctc tcc acc aac atg gct ctc tcc atc att cac aac ggt tac gct	4079
Arg Leu Ser Thr Asn Met Ala Leu Ser Ile Ile His Asn Gly Tyr Ala	
1345 1350 1355	
ggc acc agg cgc atc caa tgc aac ctc atg aag caa tac gct tcc ctt	4127
Gly Thr Arg Arg Ile Gln Cys Asn Leu Met Lys Gln Tyr Ala Ser Leu	
1360 1365 1370 1375	
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Gly Asp Lys Phe Ile Ile Tyr Asp Ser Ser Phe Asp Asp Ala Asn Arg	
1380 1385 1390	
ttc aac ttg gtc cca ctc ttc aag ttc ggc aag gat gag aac tct gat	4223

Phe Asn Leu Val Pro Leu Phe Lys Phe Gly Lys Asp Glu Asn Ser Asp	
1395 1400 1405	
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Asp Ser Ile Cys Ile Tyr Asn Glu Asn Pro Ser Ser Glu Asp Lys Lys	
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Trp Tyr Phe Ser Ser Lys Asp Asp Asn Lys Thr Ala Asp Tyr Asn Gly	
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Gly Thr Gln Cys Ile Asp Ala Gly Thr Ser Asn Lys Asp Phe Tyr Tyr	
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Asn Leu Gln Glu Ile Glu Val Ile Ser Val Thr Gly Gly Tyr Trp Ser	
1460 1465 1470	
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Ser Tyr Lys Ile Ser Asn Pro Ile Asn Ile Asn Thr Gly Ile Asp Ser	
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gcc aag gtc aag gtc act gtc aag gct ggt ggc gat gac caa atc ttc	4511
Ala Lys Val Lys Val Thr Val Lys Ala Gly Gly Asp Asp Gln Ile Phe	
1490 1495 1500	
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Thr Ala Asp Asn Ser Thr Tyr Val Pro Gln Gln Pro Ala Pro Ser Phe	
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gag gag atg atc tac caa ttc aac aac ctc acc att gac tgc aag aac	4607
Glu Glu Met Ile Tyr Gln Phe Asn Asn Leu Thr Ile Asp Cys Lys Asn	
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ctc aac ttc att gac aac cag gct cac att gag att gac ttc act gcc	4655
Leu Asn Phe Ile Asp Asn Gln Ala His Ile Glu Ile Asp Phe Thr Ala	
1540 1545 1550	
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Thr Ala Gln Asp Gly Arg Phe Leu Gly Ala Glu Thr Phe Ile Ile Pro	
1555 1560 1565	
gtc acc aag aag gtc ctt ggc act gag aac gtc att gct ctc tac tct	4751
Val Thr Lys Lys Val Leu Gly Thr Glu Asn Val Ile Ala Leu Tyr Ser	
1570 1575 1580	
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Glu Asn Asn Gly Val Gln Tyr Met Gln Ile Gly Ala Tyr Arg Thr Arg	
1585 1590 1595	
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Leu Asn Thr Leu Phe Ala Gln Gln Leu Val Ser Arg Ala Asn Arg Gly	
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Ile Asp Ala Val Leu Ser Met Glu Thr Gln Asn Ile Gln Glu Pro Gln	
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ctt ggt gct ggc acc tac gtc caa ctt gtc ttg gac aag tac gat gag	4943
Leu Gly Ala Gly Thr Tyr Val Gln Leu Val Leu Asp Lys Tyr Asp Glu	

1635	1640	1645	
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ttc aag gag aac gac tcc ttc gtc atc tac caa ggt gag ttg tct gag Phe Lys Glu Asn Asp Ser Phe Val Ile Tyr Gln Gly Glu Leu Ser Glu 1665 1670 1675			5039
acc tcc caa act gtg gtc aag gtc ttc ctc tcc tac ttc att gag gcc Thr Ser Gln Thr Val Val Lys Val Phe Leu Ser Tyr Phe Ile Glu Ala 1680 1685 1690 1695			5087
acc ggt aac aag aac cac ctc tgg gtc agg gcc aag tac cag aag gag Thr Gly Asn Lys Asn His Leu Trp Val Arg Ala Lys Tyr Gln Lys Glu 1700 1705 1710			5135
acc act gac aag atc ctc ttc gac agg act gat gag aag gac cca cat Thr Thr Asp Lys Ile Leu Phe Asp Arg Thr Asp Glu Lys Asp Pro His 1715 1720 1725			5183
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gct caa gct ctc aag aac gac tct gag cca atg gac ttc tct ggt gcc Ala Gln Ala Leu Lys Asn Asp Ser Glu Pro Met Asp Phe Ser Gly Ala 1745 1750 1755			5279
aac gct ctc tac ttc tgg gag ttg ttc tac tac act cca atg atg atg Asn Ala Leu Tyr Phe Trp Glu Leu Phe Tyr Thr Pro Met Met Met 1760 1765 1770 1775			5327
gct cac agg ctc ctt caa gag cag aac ttc gat gct gcc aac cac tgg Ala His Arg Leu Leu Gln Glu Gln Asn Phe Asp Ala Ala Asn His Trp 1780 1785 1790			5375
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gcc atc tac cac tgg aac gtc agg cca ttg gag gag gac acc tcc tgg Ala Ile Tyr His Trp Asn Val Arg Pro Leu Glu Glu Asp Thr Ser Trp 1810 1815 1820			5471
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gac cca atg cac tac aag gtg gcc acc ttc atg gcc acc ttg gac ctt Asp Pro Met His Tyr Lys Val Ala Thr Phe Met Ala Thr Leu Asp Leu 1840 1845 1850 1855			5567
ctc atg gcc aga ggt gat gct gcc tac cgc caa ttg gag agg gac acc Leu Met Ala Arg Gly Asp Ala Ala Tyr Arg Gln Leu Glu Arg Asp Thr 1860 1865 1870			5615
ttg gct gag gcc aag atg tgg tac acc caa gct ctc aac ttg ctg ggt Leu Ala Glu Ala Lys Met Trp Tyr Thr Gln Ala Leu Asn Leu Leu Gly 1875 1880 1885			5663

gat gag cca caa gtc atg ctc tcc aca acc tgg gcc aac cca acc ttg Asp Glu Pro Gln Val Met Leu Ser Thr Thr Trp Ala Asn Pro Thr Leu 1890 1895 1900	5711
ggc aac gct gcc tcc aag acc aca caa cag gtc agg caa cag gtc ctc Gly Asn Ala Ala Ser Lys Thr Thr Gln Gln Val Arg Gln Gln Val Leu 1905 1910 1915	5759
acc caa ctc agg ctc aac tct aga gtc aag act cca ctc ttg ggc act Thr Gln Leu Arg Leu Asn Ser Arg Val Lys Thr Pro Leu Leu Gly Thr 1920 1925 1930 1935	5807
gcc aac tcc ctc act gct ctc ttc ctc cca caa gag aac tcc aaa ctt Ala Asn Ser Leu Thr Ala Leu Phe Leu Pro Gln Glu Asn Ser Lys Leu 1940 1945 1950	5855
aag ggt tac tgg agg acc ctt gct caa cgc atg ttc aac ctc agg cac Lys Gly Tyr Trp Arg Thr Leu Ala Gln Arg Met Phe Asn Leu Arg His 1955 1960 1965	5903
aac ctc tcc att gat ggt caa cca ctc tcc ttg cca ctc tac gct aag Asn Leu Ser Ile Asp Gly Gln Pro Leu Ser Leu Pro Leu Tyr Ala Lys 1970 1975 1980	5951
cca gct gac cca aag gct ctc ctt tcc gct gct gtc tcc gca tcc caa Pro Ala Asp Pro Lys Ala Leu Leu Ser Ala Ala Val Ser Ala Ser Gln 1985 1990 1995	5999
ggt ggt gct gac ctc cca aag gct cca ctc acc atc cac agg ttc cca Gly Gly Ala Asp Leu Pro Lys Ala Pro Leu Thr Ile His Arg Phe Pro 2000 2005 2010 2015	6047
caa atg ttg gag ggt gcc cgt ggt ctt gtc aac cag ctc atc caa ttc Gln Met Leu Glu Gly Ala Arg Gly Leu Val Asn Gln Leu Ile Gln Phe 2020 2025 2030	6095
ggt tcc tct ctc ctt ggt tac tct gag agg caa gat gct gag gcc atg Gly Ser Ser Leu Leu Gly Tyr Ser Glu Arg Gln Asp Ala Glu Ala Met 2035 2040 2045	6143
tcc caa ctc ttg caa acc cag gct tct gag ttg atc ctc acc tcc atc Ser Gln Leu Leu Gln Thr Gln Ala Ser Glu Leu Ile Leu Thr Ser Ile 2050 2055 2060	6191
agg atg caa gac aac cag ctt gct gag ttg gac tct gag aag act gct Arg Met Gln Asp Asn Gln Leu Ala Glu Leu Asp Ser Glu Lys Thr Ala 2065 2070 2075	6239
ctc caa gtc tcc ctt gct ggt gtc caa cag agg ttc gac agc tac tcc Leu Gln Val Ser Leu Ala Gly Val Gln Gln Arg Phe Asp Ser Tyr Ser 2080 2085 2090 2095	6287
caa ctc tac gag gag aac atc aac gct ggt gag caa agg gct ttg gct Gln Leu Tyr Glu Glu Asn Ile Asn Ala Gly Glu Gln Arg Ala Leu Ala 2100 2105 2110	6335
ctc agg tct gag tct gcc att gag tcc caa ggt gct caa atc tcc cgc Leu Arg Ser Glu Ser Ala Ile Glu Ser Gln Gly Ala Gln Ile Ser Arg 2115 2120 2125	6383

atg gct ggt gct ggc gtg gac atg gct cca aac atc ttc ggt ctt gct Met Ala Gly Ala Gly Val Asp Met Ala Pro Asn Ile Phe Gly Leu Ala 2130 2135 2140	6431
gat ggt ggc atg cac tac ggt gcc att gct tac gcc att gct gat ggc Asp Gly Gly Met His Tyr Gly Ala Ile Ala Tyr Ala Ile Ala Asp Gly 2145 2150 2155	6479
att gag ctt tct gct tct gcc aag atg gtt gat gct gag aag gtg gct Ile Glu Leu Ser Ala Ser Ala Lys Met Val Asp Ala Glu Lys Val Ala 2160 2165 2170 2175	6527
caa tct gaa atc tac cgt cgc aga cgc caa gaa tgg aag atc caa agg Gln Ser Glu Ile Tyr Arg Arg Arg Arg Gln Glu Trp Lys Ile Gln Arg 2180 2185 2190	6575
gac aac gct caa gct gag atc aac cag ctc aac gct caa ctt gag tcc Asp Asn Ala Gln Ala Glu Ile Asn Gln Leu Asn Ala Gln Leu Glu Ser 2195 2200 2205	6623
ctc agc atc agg cgt gag gct gct gag atg cag aag gag tac ctc aag Leu Ser Ile Arg Arg Glu Ala Ala Glu Met Gln Lys Glu Tyr Leu Lys 2210 2215 2220	6671
acc caa cag gct caa gct cag gct caa ctc acc ttc ctc agg tcc aag Thr Gln Gln Ala Gln Ala Gln Ala Gln Leu Thr Phe Leu Arg Ser Lys 2225 2230 2235	6719
ttc tcc aac cag gct ctc tac tcc tgg ctc aga ggc cgc ctc tct ggc Phe Ser Asn Gln Ala Leu Tyr Ser Trp Leu Arg Gly Arg Leu Ser Gly 2240 2245 2250 2255	6767
atc tac ttc caa ttc tac gac ttg gct gtc tcc cgc tgc ctc atg gct Ile Tyr Phe Gln Phe Tyr Asp Leu Ala Val Ser Arg Cys Leu Met Ala 2260 2265 2270	6815
gag caa tcc tac caa tgg gag gcc aac gac aac agc atc tcc ttc gtc Glu Gln Ser Tyr Gln Trp Glu Ala Asn Asp Asn Ser Ile Ser Phe Val 2275 2280 2285	6863
aag cca ggt gct tgg caa ggc acc tac gct ggt ctc ctt tgc ggt gag Lys Pro Gly Ala Trp Gln Gly Thr Tyr Ala Gly Leu Leu Cys Gly Glu 2290 2295 2300	6911
gct ctc atc cag aac ttg gct caa atg gag gag gct tac ctc aag tgg Ala Leu Ile Gln Asn Leu Ala Gln Met Glu Glu Ala Tyr Leu Lys Trp 2305 2310 2315	6959
gag tcc aga gct ttg gag gta gag agg act gtc tcc ctt gct gta gtc Glu Ser Arg Ala Leu Glu Val Glu Arg Thr Val Ser Leu Ala Val Val 2320 2325 2330 2335	7007
tac gac tcc ttg gag ggc aac gac agg ttc aac ctt gct gag caa atc Tyr Asp Ser Leu Glu Gly Asn Asp Arg Phe Asn Leu Ala Glu Gln Ile 2340 2345 2350	7055
cca gct ctc ttg gac aag ggt gag ggc act gct ggc acc aag gag aac Pro Ala Leu Leu Asp Lys Gly Glu Gly Thr Ala Gly Thr Lys Glu Asn 2355 2360 2365	7103
ggt ctc tcc ttg gcc aac gcc atc ctc tct gct tct gtc aag ctc tct	7151

Gly Leu Ser Leu Ala Asn Ala Ile Leu Ser Ala Ser Val Lys Leu Ser  
 2370 2375 2380  
 gac ctc aag ttg ggt act gac tac cca gac tcc att gtg ggt tcc aac 7199  
 Asp Leu Lys Leu Gly Thr Asp Tyr Pro Asp Ser Ile Val Gly Ser Asn  
 2385 2390 2395  
 aag gtc aga agg atc aag caa atc tct gtc tcc ctc cca gct ttg gtg 7247  
 Lys Val Arg Arg Ile Lys Gln Ile Ser Val Ser Leu Pro Ala Leu Val  
 2400 2405 2410 2415  
 ggt cca tac caa gat gtc caa gcc atg ctc tcc tac ggt ggc tcc acc 7295  
 Gly Pro Tyr Gln Asp Val Gln Ala Met Leu Ser Tyr Gly Gly Ser Thr  
 2420 2425 2430  
 caa ctc cca aag ggt tgc tct gct ttg gct gtc tcc cac ggc acc aac 7343  
 Gln Leu Pro Lys Gly Cys Ser Ala Leu Ala Val Ser His Gly Thr Asn  
 2435 2440 2445  
 gac tct ggt caa ttc caa ctt gac ttc aac gat ggc aag tac ctc cca 7391  
 Asp Ser Gly Gln Phe Gln Leu Asp Phe Asn Asp Gly Lys Tyr Leu Pro  
 2450 2455 2460  
 ttc gaa ggc att gct ttg gat gac caa ggc acc ctc aac ctc caa ttc 7439  
 Phe Glu Gly Ile Ala Leu Asp Asp Gln Gly Thr Leu Asn Leu Gln Phe  
 2465 2470 2475  
 cca aac gcc act gac aag cag aag gcc atc ctc caa acc atg tct gac 7487  
 Pro Asn Ala Thr Asp Lys Gln Lys Ala Ile Leu Gln Thr Met Ser Asp  
 2480 2485 2490 2495  
 atc atc ctc cac atc agg tac acc atc agg tgagctcgag aggcctgcgg 7537  
 Ile Ile Leu His Ile Arg Tyr Thr Ile Arg  
 2500 2505  
 ccgc 7541

<210> 5  
 <211> 63  
 <212> DNA  
 <213> Artificial Sequence

<220>  
 <223> Description of Artificial Sequence:hemicot sequence  
 encoding ER signal from 15 kDa zein from Black  
 Mexican Sweet maize

<220>  
 <221> CDS  
 <222> (1)..(63)

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 1 5 10 15  
 gcc tgt gct tca gcc 63  
 Ala Cys Ala Ser Ala  
 20



<210> 6  
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 <212> DNA  
 <213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:hemicot tcdA  
 fused to the modified 15 kDa zein endoplasmic  
 reticulum signal peptide

<220>

<221> CDS

<222> (4)..(7614)

<400> 6

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gct gcc tgt gct tca gcc atg aac gag tcc gtc aag gag atc cca gac	96
Ala Ala Cys Ala Ser Ala Met Asn Glu Ser Val Lys Glu Ile Pro Asp	
20 25 30	
gtc ctc aag tcc caa tgc ggt ttc aac tgc ctc act gac atc tcc cac	144
Val Leu Lys Ser Gln Cys Gly Phe Asn Cys Leu Thr Asp Ile Ser His	
35 40 45	
agc tcc ttc aac gag ttc aga caa caa gtc tct gag cac ctc tcc tgg	192
Ser Ser Phe Asn Glu Phe Arg Gln Gln Val Ser Glu His Leu Ser Trp	
50 55 60	
tcc gag acc cat gac ctc tac cat gac gct cag caa gct cag aag gac	240
Ser Glu Thr His Asp Leu Tyr His Asp Ala Gln Gln Ala Gln Lys Asp	
65 70 75	
aac agg ctc tac gag gct agg atc ctc aag agg gct aac cca caa ctc	288
Asn Arg Leu Tyr Glu Ala Arg Ile Leu Lys Arg Ala Asn Pro Gln Leu	
80 85 90 95	
cag aac gct gtc cac ctc gcc atc ttg gct cca aac gct gag ttg att	336
Gln Asn Ala Val His Leu Ala Ile Leu Ala Pro Asn Ala Glu Leu Ile	
100 105 110	
ggt tac aac aac cag ttc tct ggc aga gct agc cag tac gtg gct cct	384
Gly Tyr Asn Asn Gln Phe Ser Gly Arg Ala Ser Gln Tyr Val Ala Pro	
115 120 125	
ggt aca gtc tcc tcc atg ttc agc cca gcc gct tac ctc act gag ttg	432
Gly Thr Val Ser Ser Met Phe Ser Pro Ala Ala Tyr Leu Thr Glu Leu	
130 135 140	
tac cgc gag gct agg aac ctt cat gct tct gac tcc gtc tac tac ttg	480
Tyr Arg Glu Ala Arg Asn Leu His Ala Ser Asp Ser Val Tyr Tyr Leu	
145 150 155	
gac aca cgc aga cca gac ctc aag agc atg gcc ctc agc caa cag aac	528
Asp Thr Arg Arg Pro Asp Leu Lys Ser Met Ala Leu Ser Gln Gln Asn	
160 165 170 175	
atg gac att gag ttg tcc acc ctc tcc ttg agc aac gag ctt ctc ttg	576

Met	Asp	Ile	Glu	Leu	Ser	Thr	Leu	Ser	Leu	Ser	Asn	Glu	Leu	Leu	Leu		
				180					185					190			
gag tcc atc aag act gag agc aag ttg gag aac tac acc aag gtc atg 624																	
Glu	Ser	Ile	Lys	Thr	Glu	Ser	Lys	Leu	Glu	Asn	Tyr	Thr	Lys	Val	Met		
			195					200					205				
gag atg ctc tcc acc ttc aga cca agc ggt gca act cca tac cat gat 672																	
Glu	Met	Leu	Ser	Thr	Phe	Arg	Pro	Ser	Gly	Ala	Thr	Pro	Tyr	His	Asp		
		210					215					220					
gcc tac gag aac gtc agg gag gtc atc caa ctt caa gac cct ggt ctt 720																	
Ala	Tyr	Glu	Asn	Val	Arg	Glu	Val	Ile	Gln	Leu	Gln	Asp	Pro	Gly	Leu		
	225					230					235						
gag caa ctc aac gct tct cca gcc att gct ggt ttg atg cac cag gca 768																	
Glu	Gln	Leu	Asn	Ala	Ser	Pro	Ala	Ile	Ala	Gly	Leu	Met	His	Gln	Ala		
	240				245					250					255		
tcc ttg ctc ggt atc aac gcc tcc atc tct cct gag ttg ttc aac atc 816																	
Ser	Leu	Leu	Gly	Ile	Asn	Ala	Ser	Ile	Ser	Pro	Glu	Leu	Phe	Asn	Ile		
				260					265					270			
ttg act gag gag atc act gag ggc aac gct gag gag ttg tac aag aag 864																	
Leu	Thr	Glu	Glu	Ile	Thr	Glu	Gly	Asn	Ala	Glu	Glu	Leu	Tyr	Lys	Lys		
			275					280					285				
aac ttc ggc aac att gag cca gcc tct ctt gca atg cct gag tac ctc 912																	
Asn	Phe	Gly	Asn	Ile	Glu	Pro	Ala	Ser	Leu	Ala	Met	Pro	Glu	Tyr	Leu		
		290					295					300					
aag agg tac tac aac ttg tct gat gag gag ctt tct caa ttc att ggc 960																	
Lys	Arg	Tyr	Tyr	Asn	Leu	Ser	Asp	Glu	Glu	Leu	Ser	Gln	Phe	Ile	Gly		
	305					310					315						
aag gct tcc aac ttc ggt caa cag gag tac agc aac aac cag ctc atc 1008																	
Lys	Ala	Ser	Asn	Phe	Gly	Gln	Gln	Glu	Tyr	Ser	Asn	Asn	Gln	Leu	Ile		
	320				325					330					335		
act cca gtt gtg aac tcc tct gat ggc act gtg aag gtc tac cgc atc 1056																	
Thr	Pro	Val	Val	Asn	Ser	Ser	Asp	Gly	Thr	Val	Lys	Val	Tyr	Arg	Ile		
				340					345					350			
aca cgt gag tac acc aca aac gcc tac caa atg gat gtt gag ttg ttc 1104																	
Thr	Arg	Glu	Tyr	Thr	Thr	Asn	Ala	Tyr	Gln	Met	Asp	Val	Glu	Leu	Phe		
			355				360					365					
cca ttc ggt ggt gag aac tac aga ctt gac tac aag ttc aag aac ttc 1152																	
Pro	Phe	Gly	Gly	Glu	Asn	Tyr	Arg	Leu	Asp	Tyr	Lys	Phe	Lys	Asn	Phe		
		370					375					380					
tac aac gcc tcc tac ctc tcc atc aag ttg aac gac aag agg gag ctt 1200																	
Tyr	Asn	Ala	Ser	Tyr	Leu	Ser	Ile	Lys	Leu	Asn	Asp	Lys	Arg	Glu	Leu		
	385					390					395						
gtc agg act gag ggt gct cct caa gtg aac att gag tac tct gcc aac 1248																	
Val	Arg	Thr	Glu	Gly	Ala	Pro	Gln	Val	Asn	Ile	Glu	Tyr	Ser	Ala	Asn		
	400				405					410					415		
atc acc ctc aac aca gct gac atc tct caa cca ttc gag att ggt ttg 1296																	
Ile	Thr	Leu	Asn	Thr	Ala	Asp	Ile	Ser	Gln	Pro	Phe	Glu	Ile	Gly	Leu		

46

ctc ttc atc atg acc agc acc tcc tac aac aag acc ctc act cct gag	2064
Leu Phe Ile Met Thr Ser Thr Ser Tyr Asn Lys Thr Leu Thr Pro Glu	
675 680 685	
atc aag aac ctc ttg gac aca gtc tac cac ggt ctc caa ggc ttc gac	2112
Ile Lys Asn Leu Leu Asp Thr Val Tyr His Gly Leu Gln Gly Phe Asp	
690 695 700	
aag gac aag gct gac ttg ctt cat gtc atg gct ccc tac att gca gcc	2160
Lys Asp Lys Ala Asp Leu Leu His Val Met Ala Pro Tyr Ile Ala Ala	
705 710 715	
acc ctc caa ctc tcc tct gag aac gtg gct cac tct gtc ttg ctc tgg	2208
Thr Leu Gln Leu Ser Ser Glu Asn Val Ala His Ser Val Leu Leu Trp	
720 725 730 735	
gct gac aag ctc caa cct ggt gat ggt gcc atg act gct gag aag ttc	2256
Ala Asp Lys Leu Gln Pro Gly Asp Gly Ala Met Thr Ala Glu Lys Phe	
740 745 750	
tgg gac tgg ctc aac acc aag tac aca cca ggc tcc tct gag gct gtt	2304
Trp Asp Trp Leu Asn Thr Lys Tyr Thr Pro Gly Ser Ser Glu Ala Val	
755 760 765	
gag act caa gag cac att gtg caa tac tgc cag gct ctt gca cag ttg	2352
Glu Thr Gln Glu His Ile Val Gln Tyr Cys Gln Ala Leu Ala Gln Leu	
770 775 780	
gag atg gtc tac cac tcc act ggc atc aac gag aac gct ttc aga ctc	2400
Glu Met Val Tyr His Ser Thr Gly Ile Asn Glu Asn Ala Phe Arg Leu	
785 790 795	
ttc gtc acc aag cct gag atg ttc ggt gct gcc aca ggt gct gca cct	2448
Phe Val Thr Lys Pro Glu Met Phe Gly Ala Ala Thr Gly Ala Ala Pro	
800 805 810 815	
gct cat gat gct ctc tcc ctc atc atg ttg acc agg ttc gct gac tgg	2496
Ala His Asp Ala Leu Ser Leu Ile Met Leu Thr Arg Phe Ala Asp Trp	
820 825 830	
gtc aac gct ctt ggt gag aag gct tcc tct gtc ttg gct gcc ttc gag	2544
Val Asn Ala Leu Gly Glu Lys Ala Ser Ser Val Leu Ala Ala Phe Glu	
835 840 845	
gcc aac tcc ctc act gct gag caa ctt gct gat gcc atg aac ctt gat	2592
Ala Asn Ser Leu Thr Ala Glu Gln Leu Ala Asp Ala Met Asn Leu Asp	
850 855 860	
gcc aac ctc ttg ctc caa gct tcc att caa gct cag aac cac caa cac	2640
Ala Asn Leu Leu Leu Gln Ala Ser Ile Gln Ala Gln Asn His Gln His	
865 870 875	
ctc cca cct gtc act cca gag aac gct ttc tcc tgc tgg acc tcc atc	2688
Leu Pro Pro Val Thr Pro Glu Asn Ala Phe Ser Cys Trp Thr Ser Ile	
880 885 890 895	
aac acc atc ctc caa tgg gtc aac gtg gct cag caa ctc aac gtg gct	2736
Asn Thr Ile Leu Gln Trp Val Asn Val Ala Gln Gln Leu Asn Val Ala	
900 905 910	

cca caa ggt gtc tct gct ttg gtc ggt ctt gac tac atc cag tcc atg	2784
Pro Gln Gly Val Ser Ala Leu Val Gly Leu Asp Tyr Ile Gln Ser Met	
915 920 925	
aag gag aca cca acc tac gct caa tgg gag aac gca gct ggt gtc ttg	2832
Lys Glu Thr Pro Thr Tyr Ala Gln Trp Glu Asn Ala Ala Gly Val Leu	
930 935 940	
act gct ggt ctc aac tcc caa cag gcc aac acc ctc cat gct ttc ttg	2880
Thr Ala Gly Leu Asn Ser Gln Gln Ala Asn Thr Leu His Ala Phe Leu	
945 950 955	
gat gag tct cgc tct gct gcc ctc tcc acc tac tac atc agg caa gtc	2928
Asp Glu Ser Arg Ser Ala Ala Leu Ser Thr Tyr Tyr Ile Arg Gln Val	
960 965 970 975	
gcc aag gca gct gct gcc atc aag tct cgc gat gac ctc tac caa tac	2976
Ala Lys Ala Ala Ala Ala Ile Lys Ser Arg Asp Asp Leu Tyr Gln Tyr	
980 985 990	
ctc ctc att gac aac cag gtc tct gct gcc atc aag acc acc agg atc	3024
Leu Leu Ile Asp Asn Gln Val Ser Ala Ala Ile Lys Thr Thr Arg Ile	
995 1000 1005	
gct gag gcc atc gct tcc atc caa ctc tac gtc aac cgc gct ctt gag	3072
Ala Glu Ala Ile Ala Ser Ile Gln Leu Tyr Val Asn Arg Ala Leu Glu	
1010 1015 1020	
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Asn Val Glu Glu Asn Ala Asn Ser Gly Val Ile Ser Arg Gln Phe Phe	
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atc gac tgg gac aag tac aac aag agg tac tcc acc tgg gct ggt gtc	3168
Ile Asp Trp Asp Lys Tyr Asn Lys Arg Tyr Ser Thr Trp Ala Gly Val	
1040 1045 1050 1055	
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Ser Gln Leu Val Tyr Tyr Pro Glu Asn Tyr Ile Asp Pro Thr Met Arg	
1060 1065 1070	
att ggt cag acc aag atg atg gat gct ctc ttg caa tct gtc tcc caa	3264
Ile Gly Gln Thr Lys Met Met Asp Ala Leu Leu Gln Ser Val Ser Gln	
1075 1080 1085	
agc caa ctc aac gct gac act gtg gag gat gcc ttc atg agc tac ctc	3312
Ser Gln Leu Asn Ala Asp Thr Val Glu Asp Ala Phe Met Ser Tyr Leu	
1090 1095 1100	
acc tcc ttc gag caa gtt gcc aac ctc aag gtc atc tct gct tac cat	3360
Thr Ser Phe Glu Gln Val Ala Asn Leu Lys Val Ile Ser Ala Tyr His	
1105 1110 1115	
gac aac atc aac aac gac caa ggt ctc acc tac ttc att ggt ctc tct	3408
Asp Asn Ile Asn Asn Asp Gln Gly Leu Thr Tyr Phe Ile Gly Leu Ser	
1120 1125 1130 1135	
gag act gat gct ggt gag tac tac tgg aga tcc gtg gac cac agc aag	3456
Glu Thr Asp Ala Gly Glu Tyr Tyr Trp Arg Ser Val Asp His Ser Lys	
1140 1145 1150	
ttc aac gat ggc aag ttc gct gca aac gct tgg tct gag tgg cac aag	3504

Phe Asn Asp Gly Lys Phe Ala Ala Asn Ala Trp Ser Glu Trp His Lys  
 1155 1160 1165

att gac tgc cct atc aac cca tac aag tcc acc atc aga cct gtc atc 3552  
 Ile Asp Cys Pro Ile Asn Pro Tyr Lys Ser Thr Ile Arg Pro Val Ile  
 1170 1175 1180

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 1185 1190 1195

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 Lys Gln Thr Gly Asn Ser Lys Asp Gly Tyr Gln Thr Glu Thr Asp Tyr  
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 Arg Tyr Glu Leu Lys Leu Ala His Ile Arg Tyr Asp Gly Thr Trp Asn  
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 Thr Pro Ile Thr Phe Asp Val Asn Lys Lys Ile Ser Glu Leu Lys Leu  
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 1250 1255 1260

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 Asp Thr Leu Leu Val Met Phe Tyr Asn Gln Gln Asp Thr Leu Asp Ser  
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 Tyr Lys Asn Ala Ser Met Gln Gly Leu Tyr Ile Phe Ala Asp Met Ala  
 1280 1285 1290 1295

tcc aag gac atg act cca gag caa agc aac gtc tac cgt gac aac tcc 3936  
 Ser Lys Asp Met Thr Pro Glu Gln Ser Asn Val Tyr Arg Asp Asn Ser  
 1300 1305 1310

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acc atc aac tac aag gct gcc tct tcc gac ctc aaa atc tac atc agc 4128  
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 1360 1365 1370 1375

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 Pro Lys Leu Arg Ile Ile His Asn Gly Tyr Glu Gly Gln Lys Arg Asn  
 1380 1385 1390

cag tgc aac ttg atg aac aag tac ggc aag ttg ggt gac aag ttc att 4224  
 Gln Cys Asn Leu Met Asn Lys Tyr Gly Lys Leu Gly Asp Lys Phe Ile

1395	1400	1405	
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atg ttc tac cca gtc tac caa tac tct ggc aac acc tct ggt ctc aac Met Phe Tyr Pro Val Tyr Gln Tyr Ser Gly Asn Thr Ser Gly Leu Asn 1425 1430 1435			4320
cag ggt aga ctc ttg ttc cac agg gac acc acc tac cca agc aag gtg Gln Gly Arg Leu Leu Phe His Arg Asp Thr Thr Tyr Pro Ser Lys Val 1440 1445 1450 1455			4368
gag gct tgg att cct ggt gcc aag agg tcc ctc acc aac cag aac gct Glu Ala Trp Ile Pro Gly Ala Lys Arg Ser Leu Thr Asn Gln Asn Ala 1460 1465 1470			4416
gcc att ggt gat gac tac gcc aca gac tcc ctc aac aag cct gat gac Ala Ile Gly Asp Asp Tyr Ala Thr Asp Ser Leu Asn Lys Pro Asp Asp 1475 1480 1485			4464
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gtc tct ggt cca gtg gag atc aac act gca atc agc cca gcc aag gtc Val Ser Gly Pro Val Glu Ile Asn Thr Ala Ile Ser Pro Ala Lys Val 1505 1510 1515			4560
caa atc att gtc aag gct ggt ggc aag gag caa acc ttc aca gct gac Gln Ile Ile Val Lys Ala Gly Gly Lys Glu Gln Thr Phe Thr Ala Asp 1520 1525 1530 1535			4608
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Pro His Phe Val Arg Asp Asp Lys Gly Ile Val Thr Ile Asn Pro Lys	
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Val Asp Pro Asp Ala Val Ala Gln His Asp Pro Met His Tyr Lys Val	
1860 1865 1870	
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Ser Thr Phe Met Arg Thr Leu Asp Leu Leu Ile Ala Arg Gly Asp His	
1875 1880 1885	



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2160 2165 2170 2175	
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2275 2280 2285	
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2290 2295 2300	
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2320 2325 2330 2335	
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2340 2345 2350	
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Val Glu Arg Thr Val Ser Leu Ala Glu Val Tyr Ala Gly Leu Pro Lys	
2355 2360 2365	
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2385	2390	2395	
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2420	2425	2430	
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2435	2440	2445	
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2450	2455	2460	
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Asn Gly Cys Glu Ala Leu Ala Val Ser His Gly Met Asn Asp Ser Gly			
2465	2470	2475	
caa ttc caa ctt gac ttc aac gat ggc aag ttc ctc cca ttc gag ggc			7488
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2480	2485	2490	2495
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Ile Ala Ile Asp Gln Gly Thr Leu Thr Leu Ser Phe Pro Asn Ala Ser			
2500	2505	2510	
atg cca gag aag gga aag caa gcc acc atg ctc aag acc ctc aac gat			7584
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2515	2520	2525	
atc atc ctc cac atc agg tac acc atc aag tgagctc			7621
Ile Ile Leu His Ile Arg Tyr Thr Ile Lys			
2530	2535		

## INTERNATIONAL SEARCH REPORT

Internat. Application No  
PCT/US 00/22237

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N9/52 C12N15/82 C07K14/24 C12N15/11

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

STRAND, EPO-Internal, WPI Data, PAJ

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 08932 A (DOW AGROSCIENCES LLC ; WISCONSIN ALUMNI RES FOUND (US)) 5 March 1998 (1998-03-05) cited in the application SEQ ID NO:11 in this document is the unmodified version of SEQ ID NO:3 of the present application. SEQ ID NO:46 corresponds to SEQ ID NO:5. page 16, line 31 -page 19, line 35	1-7
A	WO 97 13402 A (DOWELANCO) 17 April 1997 (1997-04-17) the whole document	1-7

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Date of the actual completion of the international search

1 December 2000

Date of mailing of the international search report

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Information on patent family members

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